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Supplemental Data

XLID-Causing Mutations and Associated Genes Challenged

in Light of Data From Large-Scale Human Exome Sequencing

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ZNF81 (zinc finger protein 81): questionable

An (X;9) *de novo* translocation interrupting *ZNF81* on the X-chromosome and *EHMT1* on chromosome 9 was identified in an intellectually disabled female. Because *ZNF81* is highly homologous to *ZNF41* (considered to be involved in ID) the disruption of *ZNF81* was considered as the cause of ID reported in the individual. In the same study, after screening 300 XLID families including 24 that mapped to the Xp11.2 locus and 217 males with ID, a single nucleotide change c.536G>A (p.Ser179Asn) fully co-segregating with the disease status in the family was detected. Although this missense mutation is not reported in EVS, one truncating variant c.1150A>T (p.Lys384*) is present in one male, while no additional *ZNF81* mutation was detected in the 208 families with suggestive XLID screened by Tarpey et *al.*² Two very recent studies described XLID probands carrying microduplications at locus Xp11.2-p11.3: one encompassing *ZNF81* and two other genes implicated in ID (*FTSJ1* and *SYN1*), the second of 335 kb including *ZNF81* with two genes non associated to cognitive disorders. The authors suggested that *ZNF81* over-expression may thus contribute to the phenotype. To summarize, the presence of a truncating variant in a male in EVS, while not decisive, weakens the proposed association of *ZNF81* to XLID, that appears excluded for its homolog ZNF41.

ATP6AP2 ($ATPase\ H+\ transporting,\ lysosomal\ accessory\ protein\ 2,\ (pro)renin\ receptor)$: questionable

The *ATP6AP2*/(pro)renin receptor gene, present in the linkage region of a large family with XLID and epilepsy was sequenced, revealing a silent change c.321C>T (p.Asp107Asp) resulting in splicing aberration in 50% of the transcripts. It was hypothesized that the resulting abnormal protein may have a dominant negative effect.⁵ No other putative mutation was observed in the 25 other genes of the candidate region. The p.Asp107Asp mutation was absent from 1,200 control X-chromosomes and is not reported in EVS. The presence of a splice variant in one male of the NHLBI population is however puzzling and might question the role of *ATP6AP2* in ID and epilepsy. Another *de novo* truncating mutation was recently reported in an abstract form, in an individual with a severe ID/epilepsy phenotype (Chitayat et al., ASHG meeting 2012, G. Nguyen personal communication). Full publication of this observation would lift the ambiguity on *ATP6AP2* involvement in ID.

ZCCHC12/SIZN1 (zinc finger CCHC domain-containing protein 12): questionable

Based on its expression in basal forebrain cholinergic neurons, *ZCCHC12/SIZN1* was considered as a good XLID candidate and was screened for mutations in a cohort of 729 ID males.⁶ Two missense mutations were identified: c.1031C>T (p.Thr344Ile) was identified in four affected male sibs in one XLID family and was absent from 494 chromosomes, and c.19C>T (p.Arg7Cys) was detected in four unrelated ID males but was also present in one control individual. This latter variant is present in EVS with a MAF of 0.3% excluding its implication in ID. Since 2008, no additional ID case has been reported with *ZCCHC12* mutations. The implication of *ZCCHC12* in ID thus remains based on the finding of one single missense variant in a single family.

IGBP1 (immunoglobulin-binding protein 1): never replicated

IGBP1 encodes the regulatory subunit α-4 of the protein phosphatase 2A (PP2A), which interacts with MID1, the product of the gene mutated in X-linked Opitz GBBB syndrome (MIM#300000) in which clinical manifestations overlap with the ones reported in the FG Syndrome (FGS). In two brothers with suspected FGS and presenting with an agenesis of the corpus callosum, ID and facial dysmorphy, sequence analysis of the FGS1 linked region identified a substitution of two adjacent nucleotides in the 5' UTR of *IGBP1* (c.-57delT and c.-55T>A), marginally affecting its expression (an observed 20% increase). Both variations were not present in 410 control chromosomes analyzed by Graham et *al.*, and since this region is not covered in EVS we were not able to check for their presence in the larger NHLBI control population. However, other 5' UTR variants located closer to the initiation ATG codon are reported in EVS. This gene, which is present in two XLID diagnostic panel lists, requires additional evidences to strengthen its implication in ID.

NLGN3 (neuroligin 3): never replicated

Neuroligins 3 and 4 both encode cell adhesion molecules present at the postsynaptic side of the synapses. Screening of both genes in 36 sib-pairs and 122 trios with ASD led to the simultaneous identification of point mutations in *NLGN4X* and *NLGN3*. The c.1411C>T; p.Arg451/471Cys missense change identified in *NLGN3* in two brothers with ASD is not present in the NHLBI population and was functionally validated. Indeed, the *Nlgn3* knock-in mouse displayed an increase in inhibitory synaptic transmission and an "autistic-like" phenotype. Talebizadeh et *al.* identified in lymphoblastoid cells an alternative transcript of *NLGN3* that lacked exon 7 and encoded a new truncated protein, present in all 30 control individuals and in all but one of the 10 ASD females tested. The authors speculated that the lack of the specific truncated isoform in this female might be implicated in her autistic phenotype. In

To conclude, contrary to *NGLN4X* for which involvement in ASD and ID has been largely replicated in independent studies, ¹²⁻¹⁴ no other mutation has been reported in *NLGN3* since 2003 in spite of a consequent number of cohorts tested (almost 700 ASD probands in total), and thus to confirm its full implication in cognitive disorders supplementary evidences are required. ¹⁵⁻²⁰

CCDC22 (coiled-coil domain-containing protein 22) : awaiting for replication

Following expression profiling in lymphoblasts of 64 XLID subjects, a five-fold decrease in the expression of *CCDC22* mRNA was observed in one male from a large family (LOD score= 2.7), who carried the missense variant c.49A>G (p.Thr17Ala) in that gene.²¹ Although the latter variation is not present in EVS, additional evidences are obviously needed to support *CCDC22* implication in ID.

CLIC2 (Chloride intracellular channel 2): awaiting for replication

The re-sequencing of the X-exome in males with putative XLID allowed the detection of a variant resulting in one missense mutation in *CLIC2*.²² This variation (c.303C>G; p.His101Gln) was present in two brothers and their mother who had a low IQ. Both males presented with severe ID, seizures, atrial fibrillation, cardiomegaly and congestive heart failure. Four other non-synonymous substitutions were reported at polymorphic frequencies in this gene. The authors suggested by an *in silico* analysis that the missense mutation p.His101Gln might have an important effect on stability, dynamics and hydrogen bond network when compared to predicted effects of four missense variants found in the control population. This missense mutation was then shown to have a functional effect, activating ryanodine receptor Ca²⁺ channel, consistent with the observed cardiac phenotype.²³ This gene needs however additional evidences to be fully considered as involved in cognitive disorders.

HCFC1 (host cell factor C1): awaiting for replication

The mutation responsible for ID in the MRX3 family remained unsolved for a long time after the initial linkage analysis of 1991. A targeted massively parallel re-sequencing of the genomic linkage interval recently allowed the identification of a regulatory point-mutation in a functional binding site for the YY1 transcription factor in the *HCFC1* gene promoter, leading to an up-regulation of its expression in lymphoblastoid cells. Screening of additional unsolved XLID families identified at least one additional missense change c.674G>A (p.Ser225Asn) segregating with the disease in the family.²⁴ Both variations are not present in EVS, and were also described in a female with schizophrenia and a

boy with autism.²⁵ However, as *HCFC1* encodes a large protein of 2,035 amino acids, the probability to identify a missense variant in this gene is high (22 missense variants predicted as possibly or probably-damaging are detected within the NHLBI cohort). Additional evidences would therefore be useful to definitely confirm the implication of *HCFC1* in ID.

NAA10 (N-alpha-acetyltranferase 10): awaiting for replication

NAA10 was very recently reported as implicated in monogenic forms of ID, and encodes the catalytic subunit of the major human N-terminal acetyltransferase. It was identified via an X-exome sequencing strategy after selection of male probands apparently sharing the same lethal X-linked pleiotropic phenotype: aged appearance, dysmorphic features, hypotonia, developmental delay, cryptorchidism and cardiac arythmias.²⁶ A unique missense mutation c.109T>C (p.Ser37Pro) that fully segregated with the disease status in both unrelated families was detected. This mutation was correlated to an in vitro reduction of 60-80% N-terminal acetylation activity for three peptides. The catalytic subunit of the NatA complex was demonstrated to be essential for survival in several organisms. Hence, one can presume that a total loss-of-function would lead to embryonic lethality in humans, and it may also explain the early death of N-terminal acetyltransferase deficient individuals (Ogden syndrome; OGDNS: MIM# 300855) carrying the partial loss-of-function p.S37P missense mutation. A second recent study screening simplex probands with non-syndromic ID and their parents through exome sequencing allowed the detection of another de novo variation c.346C>T resulting in a missense change (p.Arg116Trp) in one male.²⁷ This mutation was proposed to lead to a phenotype that does not coincide with the one described in OGDNS probands, besides the large ears, hypotonia and enlarged cerebral ventricles. All male probands affected with OGDNS manifest severe developmental delay and die before the age of two. The p.Arg116Trp missense change appears less damaging than the initial p.S37P and is in fact predicted as benign by PolyPhen. The Arg116 residue is located within the acetyltransferase domain of the protein, and the authors proposed that the change into a tryptophan might lead to an interference with Coenzyme A binding and to a reduced enzymatic activity. None of these missense mutations are reported within the NHLBI population. While the implication of NAA10 in OGDNS appears very convincing, its implication in non-syndromic ID would require additional evidence.

RPL10 (ribosomal protein L10): awaiting for replication

RPL10 was selected for reassessment of its implication in ID because only two nucleotide substitutions c.616C>A and c.639C>G have been described in males with ASD. 28: 29 It encodes a key protein for both the assembly of the large ribosomal subunit and global protein synthesis. The resulting missense changes p.Leu206Met and p.His213Gln were detected following the sequencing of candidate genes within linked regions in 296 families with ASD and with/without associated ID, and appeared to have subtle effect on ribosomal profile in a yeast complementation assay. The p.His213Gln missense mutation was later identified in an unrelated simplex case, inherited from a carrier mother. Both missense variations are located at the very C-terminal end of the protein in an eukaryote-specific extension, and appear not so conserved. They are nonetheless not observed within the NHLBI population. Additional screening of 189 individuals with ASD did not lead to the identification of new RPL10 mutations, while mutations have been reported in individuals suffering from Leukemia. Thus, evidences clearly linking the reported RPL10 missense mutations to the resulting ASD phenotype remain insufficient, but the fact that the same missense mutation (p.His123Gln) was observed in two independent families and is not reported within the NHLBI population is suggestive of an implication in cognitive disorders.

SHROOM4/KIAA1202 (shroom family member 4): awaiting for replication

After investigating the breakpoints of two balanced (X;autosome) translocations in unrelated females with moderate ID, *KIAA1202/SHROOM4* was pointed as a candidate for implication in ID due to its disruption in both cases.³² Another pathogenic CNV encompassing *SHROOM4* was detected in one XLID family.³³ Additional screening of 200 XLID families for structural variations or mutations in this gene revealed a single variation c.3266C>T (p.Ser1089Leu) in the linkage region of a large fourgeneration family that defined the Stocco dos Santos syndrome as a combination of severe ID, short stature and congenital hip luxation (MIM #300434; LOD score= 3.02).³² This mutation, absent from

over 1,000 control X-chromosomes, is not reported in EVS. A systematic re-sequencing study of the X-chromosome identified in ID individuals a recurrent in-frame insertion that was ultimately also reported in controls and six missense variants that do not seem involved in ID.² We also recently identified an inherited truncating mutation in one male with severe ID, short stature, hypotonia and dysmorphic traits (unpublished data). However, when performing segregation analyses, the variant was depicted in both unaffected brothers, therefore ruling out the pathogenicity of this variant (unpublished data). With a truncating variant observed in controls at hemizygous state, these findings do not support the implication of *SHROOM4* in cognitive disorders.

ZDHHC15, KLF8, KIAA2022, ZNF261/ZMYM3, CNKSR2 and FRMPD4

The implication of these six genes (*ZDHHC15*, *KLF8*, *KIAA2022*, *ZNF261/ZMYM3*, *CNKSR2* and *FRMPD4*) in ID relies for each of them on one single chromosomal rearrangement identified in simplex cases (**Table 2**). *ZDHHC15*, *KLF8*, *KIAA2022* and *ZNF261/ZMYM3* remain however commonly cited in XLID reviews and – with the exception of *ZNF261/ZMYM3* – are generally included in XLID diagnostic panels even though no additional cases has been published since the initial studies, more than eight years ago. Additional sequencing of *KIAA2022* and *KLF8* in 20 XLID families/20 ID male probands respectively in the initial studies failed to identify any other point mutation. More recently, duplication and deletion of two other genes, *CNKSR2* and *FRMPD4*, were described. Such findings will also need to be replicated for both genes to be fully considered as confirmed genes involved in ID.

The examination of EVS for all six genes does not bring any complementary information: no truncating variations are reported within the NHLBI cohort for any of those. Three mutations in *KIAA2022* leading to partial or total loss of its expression have been recently reported (in abstract form) to be present in five individuals with a distinctive syndromic XLID (Van Maldergem et *al.*, ASHG meeting 2012). Publication of the cognate work should unambiguously prove implication of this gene in ID and/or autism.

PTCHD1 (Patched domain-containing protein 1): likely

PTCHD1 was initially considered as a potential ASD candidate when found to be disrupted via an inherited 160 kb deletion observed in dizygotic twin brothers in a large genome-wide analysis of structural variations in ASD subjects. 40 A later similar study genotyping 996 ASD probands and over a thousand controls confirmed these findings, revealing six additional males carrying deletions disrupting PTCHD1.41 An analogous study of the implication in ID of structural variants of the Xchromosome reported a 90 kb deletion spanning entire *PTCHD1* in one XLID family. 42 Subsequently, two large mutation-screening strategies in cohorts of both ID and ASD individuals led to the identification of one additional deletion disrupting *PTCHD1* in two unrelated ID males, ^{43; 44} and seven missense variations in six ASD and two ID probands respectively (see **Table 2**). 44 Two of them (c.217C>T; p.Leu73Phe and c.1409C>A; p.Ala470Asp) did not fully segregate with the ID status in the family. Interestingly, two out of the seven missense variations are detected among the NHLBI population: the non-segregating p.Leu73Phe is reported in one heterozygous female and the c.517A>G; p.Ile173Val in two males and two females. However, since none of the other missense mutations are reported in EVS, the expression pattern in fetal and adult brain along with the cumulative evidences through CNV and point mutation analysis in both ASD and ID probands support the implication of *PTCHD1* in cognitive disorders.

SYN1 (synapsin-1): likely

SYN1 was first suggested as a candidate for Rett syndrome (MIM# 312750) due to its proposed role in the modulation of neurotransmitter release. It was later screened because it was located in the linkage region of a family with epilepsy and some learning difficulties (LOD score = 3.65) and a truncating mutation c.1197G>A (p.Trp356*) was identified, which fully segregated with the disease status. Another study supported those results, with the detection of another truncating mutation in a large family with ASD, low IQ and epilepsy, and of three other missense variations (c.152C>G; p.Ala51Gly, c.1648G>A; p.Ala550Thr, c.1699A>G; p.Thr567Ala) in additional families with similar phenotype. Interestingly, the missense mutation p.A550T was detected in four unrelated individuals

with different phenotypes: one female and one male presenting with only epilepsy, one female with both epilepsy and ASD, and one male with only ASD, hence suggesting variable phenotypic expression. The missense variant p.Ala51Gly was present in one epileptic female, and in one ASD male also carrying the p.Thr567Ala variant, already suggesting one of the two variants might not contribute to the phenotype. The latter encoded p.Ala51Gly missense variant is indeed an innocuous variant since it is reported in the NHLBI population in 11 hemizygous males and 30 carrier females. Although some doubts remain about the pathogenicity of the p.Thr567Ala variant, EVS data is not informative since the corresponding region is not covered. The *SYN1* KO mouse presents a global disorganization of synaptic vesicles at presynaptic terminals, hence suggesting an important role for formation and maintenance of presynaptic structure. He suggested human disease phenotype. He suggested human disease phenotype.

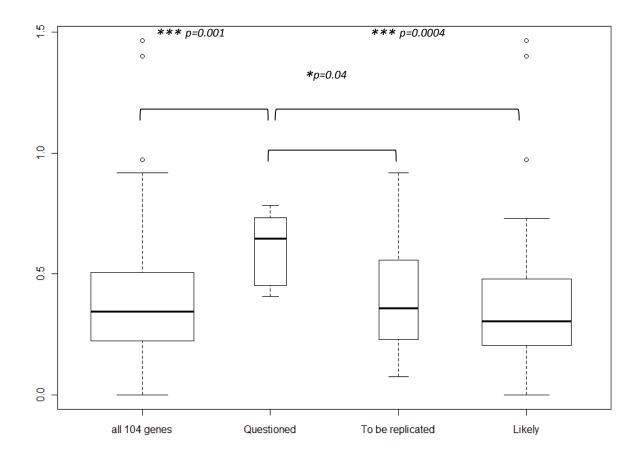


Figure S1. Distribution of the Computed dN/dS Ratios for All 104 Selected Genes and by Category, according to Their Putative Association with Intellectual Disability When Mutated

The dN/dS ratios distribution is compared between several categories: all genes, the 10 genes whose association in ID is questioned, the 15 genes which need to be replicated and the remaining genes with consistent association with monogenic forms of ID when mutated. The boxplot was generated in R: the width of all boxes are proportional to $\sqrt{\text{(number of values)}}$. Circles represent outliers ranging outside of the interval whiskers, which extend up to 1.5x (3rd quartile - 1st quartile). Significance levels between all gene subsets are computed using a Student's t-test with unequal variances in R.

Table S1. Catalog of Single Nucleotide Truncating Variants Reported among the EVS Population in All Selected Genes, Supposedly Implicated in XLID When Mutated

Gene (transcript#)	Lists	Truncating variant (p./protein length)	Truncating variant (c.)	Variant occurence (: #X-chr)	#Males (hemz)	#Females (htz)	Comments
AGTR2 (NM_000686.4)	L, R, G, E, A	p.Trp158* (/364)	c.474G>A	1:10,563	-	1 F	
ATP6AP2 (NM_005765.2)	L, R, G, E, A	splice variant (g.X:40464811)	c.859-2A>C	1:10,559	1 M	-	one isoform uses an alternative AG for this exon
ATRX (NM_000489.3)	L, R, G, E, A	p.Leu1022* (/2455)	c.3065A>C	1:10,545	-	1 F	-
BCOR (NM_001123385.1)	L, R, G, E, A	splice variant (g.X:39922860; rs147281758)	c.3847+1C>G	1:10,561	-	1 F	-
DLG3 (NM_020730.2)	L, R, G, E, A	p.Trp24* (/513)	c.71G>A	1:9,797	1 M	-	affects only isoform#2 (NM_020730.2)
		p.Trp3416* (/3686)	c.10247C>T	2:10,560	-	2 F	
DMD (NM_004006.2)	L, R, G, E	splice variant (g.X:31196048; rs145603325)	c.10262+1C>T	6:10,561	-	6 F	
(1.112_00.1000.12)		splice variant (g.X:32429867; rs147474070)	c.4233+2G>A	3:10,561	1 M	2 F	
		splice variant (g.X:32591646)	c.1812+1C>T	1:10,558	-	1 F	
MAGT1 (NM_032121.5)	L, R, G, E	p.Ser24* (/368)	c.71G>C	1:10,558	1 M	-	
NXF5 (NM_032946.2)	L, R, G,A	p.Arg320* (/366) rs140252282	c.958G>A	33:10,563 (MAF=0,31%)	7 M	26 F	-
(NWI_032940.2)		p.Cys54* (/366)	c.162G>T	1:10,563	1 M	-	-
<i>OFD1</i> (NM_003611.2)	L, R, G, E, A	splice variant (g.X:13769366)	c.936-2A>G	1:10,554	1 M	-	exon not present in all isoforms
SHROOM4 (NM_020717.3)	L, R, G, E, A	p.Trp128* (/1494) rs142159861	c.383C>T	1:10,563	-	1 F	
SLC9A6 (NM_001042537.1)	L, R, G, E, A	splice variant (g.X:135095107; rs149044510)	c.1042-1C>T	1:10,563	-	1 F	-
UPF3B (NM_080632.2)	L, R, G, E	splice variant (g.X:118974608)	c.846+1C>T	1:10,563	-	1 F	
ZNF41 (NM_007130.2)	L, R, G, E, A	splice variant (g.X:47315319; rs143323743)	c.295+1C>T	1:10,563	1 M	-	-
		p.Trp41* (/780)	c.122C>T	1:10,563	-	1 F	-
ZNF674	L, R, E, A	p.Asp442* (/582)	c.1324C>A	5:10,509	-	5 F	
(NM_001039891.2)	. , ,	p.Arg201* (/582)	c.601G>A	44:9,514 (MAF=0,46%)	19 M	25 F	
ZNF81 (NM_007137.3)	L, R, G, E, A	p.Lys384* (/662)	c.1150A>T	1:10,533	1 M	-	

XLID lists in which genes are included: L=Lubs et al.,⁵² R=Ropers et al.,⁵³ G=Greenwood, E=Emory, A=Ambry. When a variant is reported in dbSNP137, the subsequent identifier is indicated in italic (*rs#*). Gender of control individuals carrying the variant: M: Male, F: Female.

Table S2. List of XLID Mutations Reported in OMIM and in Literature for the 104 Selected Genes (*Additional Mutations from the Literature Only)

Gene	Mutation
ABCD1	, GLU291LYS
ABCD1	, PRO484ARG
ABCD1	, IVS6AS, A-G, c.1635-2, g.153006026
ABCD1	, IVS8AS, G-A, -10, 8-BP INS
ABCD1	, ARG389GLY
ABCD1	, ASN148SER
ABCD1	, TYR174ASP
ABCD1	, GLY266ARG
ABCD1	, ARG401GLN
ABCD1	, ARG418TRP
ABCD1	, ARG464TER
ABCD1	, 2-BP DEL, 1801AG
ABCD1	, GLU477TER
ABCD1	, SER515PHE
ABCD1	, 1-BP DEL, 1937C
ABCD1	, ARG518TRP
ABCD1	, IVS6DS, G-A, g.153005692, c.1634+1
ABCD1	, 2-BP DEL, 2177TA
ABCD1	, SER606LEU
ABCD1	, 1-BP DEL, 2204G
ABCD1	, ARG617HIS
ABCD1	, ARG617CYS
ABCD1	, 3-BP DEL, 1258GAG, GLU291DEL
ABCD1	, IVS8DS, G-A, c.1865+1, g.153008526
ABCD1	, IVS1DS, G-A, c.901-1, g.152994686
ABCD1	, 26-BP DEL, NT369
ACSL4,	ARG529SER
ACSL4,	IVS10, A-G, -2
ACSL4,	PRO375LEU
AFF2, (GCC)n EXPANSION
AFF2, 1	21- to 145-KB DEL
AGTR2	, GLY21VAL
AGTR2	, 1-BP DEL, 395T
AGTR2	, ARG324GLN
AGTR2	, ILE337VAL
AGTR2	, ILE53PHE
AP1S2,	GLN36TER
AP1S2,	ARG52TER
AP1S2,	4-BP DEL, NT180
AP1S2,	IVS3DS, G-A, +5
AP1S2,	GLN66TER
ARHGE	F6, IVS1AS, T-C, -11
ARHGE	F9, GLY55ALA
ARHGE	F9, GLN2TER
A DATE OF	

ARX, (GCG)10+7

ARX, 24-BP DUP, NT428

ARX, PRO353LEU

ARX, 1,517-BP DEL

ARX, 32-BP DEL, NT420

ARX, 1-BP DEL, 790C

ARX, ARG332HIS

ARX, GLN373TER

ARX, 1-BP INS, 1188C

ARX, EX1-2DEL

ARX, 1-BP DEL, 1372G

ARX, LEU343GLN

ARX, LEU33PRO

ARX, GLY286SER

ARX, THR333ASN

ARX, GLU369TER

ARX, 33-BP DUP

ARX, 24-BP DEL, NT441

ARX, 1-BP DEL, 617G

ARX, GLU78TER

ARX, 1-BP DEL, 1465G

ARX, 27-BP DUP, NT430

ARX, TYR27TER

ARX, LEU535GLN

ATP6AP2, ASP107ASP

ATP7A, IVSXDS, A-T, +3

ATP7A, IVSAS, 2642A-G, -2

ATP7A, SER637LEU

ATP7A, 8-BP DEL, NT1552

ATP7A, ARG980TER

ATP7A, IVS6DS, T-A, +6

ATP7A, IVS6DS, G-A, +1

ATP7A, 1-BP DEL, 4497G

ATP7A, GLY1019ASP

ATP7A, EX8 DEL

ATP7A, 8-BP DEL, NT408

ATP7A, EX3-4 DEL

ATP7A, ASN1304SER

ATP7A, ARG201TER

ATP7A, THR994ILE

ATP7A, PRO1386SER

ATRX, HIS750ARG

ATRX, CYS755ARG

ATRX, LYS792ASN

ATRX, ASN1002SER

ATRX, ASP1177VAL

ATRX, TYR1226HIS

ATRX, TYR1305CYS

ATRX, ARG1528TER

ATRX, GLU1530TER

ATRX, IVSAS, T-A, -10

ATRX, ARG1272GLN

ATRX, PRO852SER

ATRX, 751A-G

ATRX, PRO73ALA

ATRX, ARG129CYS

ATRX, ARG1742LYS

ATRX, IVS34, A-G, -2

ATRX, ARG246CYS

ATRX, THR1621MET

ATRX, IVS1DS, G-A, +1

ATRX, SER79TER

ATRX, ARG37TER

ATRX, LEU409SER

ATRX, ILE2052THR

ATRX, CYS220TYR

ATRX, ARG2271GLY

BCOR, PRO85LEU

BCOR, IVS8AS, G-T, -1

BCOR, ARG976TER

BCOR, 1-BP DEL, 3881A

BCOR, EX9-15DEL

BCOR, 2-BP DEL, 2488AG

BCOR, 1-BP DEL, 3286G

BCOR, 60-KB DEL

BCOR, 1-BP DEL, 1315C

BRWD3, IVS29, G-T, +1

BRWD3, 1-BP INS, 946A

BRWD3, LYS1596GLU

CASK, ARG639TER

CASK, 915G-A, EX9

CASK, ARG28LEU

CASK, TYR268HIS

CASK, ASP710GLY

CASK, TRP914ARG

CASK, PRO396SER

CASK, TYR728CYS

CASK, IVS25AS, A-T, -2

CASK, ARG106TER

CASK, 100-KB DEL

CASK, GLN547TER

CCDC22, THR17ALA

CDKL5, 1-BP DEL, 183T

CDKL5, IVSAS13, G-A, -1

CDKL5, CYS152PHE

CDKL5, ARG175SER

CDKL5, 4-BP DEL, 166GAAA

CDKL5, 2-BP DEL, 2636CT

CDKL5, GLN834TER

CDKL5, IVS6AS, G-T, -1

CDKL5, ALA40VAL

CDKL5, ILE72THR

CDKL5, THR288ILE

CDKL5, CYS291TYR

CDKL5, 2-BP INS, 903GA

CDKL5, ARG178PRO

*CLIC2, HIS101GLN

CUL4B, c.1007_1011delTTATA, p.I336fs*2

CUL4B, ARG572CYS

CUL4B, ARG388TER

*CUL4B, THR831THR

*CUL4B, R856TER

CUL4B, IVS6, A-G, -2

*CUL4B, IVS7, A-G, -2

*CUL4B, VAL745ALA

DCX, ASP62ASN

DCX, ARG192TRP

DCX, TYR125HIS

DCX, IVS4, G-A, +1

DCX, ARG59LEU

DCX, THR203ARG

DCX, SER47ARG

DCX, 2-BP INS, 36AG

DCX, 2-BP DEL, 684CT

DCX, 2-BP DEL, 691CT

DCX, ARG78HIS

DCX, ARG89GLY

DCX, ARG196HIS

DCX, ALA71SER

DKC1, PHE36VAL

DKC1, LEU37DEL

DKC1, PRO40ARG

DKC1, LEU72TYR

DKC1, GLY402GLU

DKC1, ALA353VAL

DKC1, 2-KB DEL

DKC1, -141C-G

DKC1, IVS1, C-G, +592

DKC1, THR49MET

DKC1, SER121GLY

DKC1, ILE38THR

DKC1, GLN31LYS

DKC1, THR357ALA

DKC1, IVS12DS, G-A, +1

DLG3, IVS6DS, G-A, +5

DLG3, IVS8DS, G-A, +1

DLG3, 1-BP INS, 1325C

DLG3, SER458TER

- DMD, GLU1157TER
- DMD, PROMOTER DEL
- DMD, GLU931TER
- DMD, GLN1851TER
- DMD, ARG2982TER
- DMD, IVS68, T-A, +2
- DMD, ARG3370TER
- DMD, EX73-76DEL
- DMD, 1-BP DEL, 10662T
- DMD, 1-BP INS, EX12
- DMD, AG-T, EX48
- DMD, EX21DEL
- DMD, EX18DEL
- DMD, GLN2319TER
- DMD, ARG768TER, C-T, NT2510
- DMD, 1-BP DEL, 2568C
- DMD, GLU772TER, G-T, NT2522
- DMD, IVS19, A-C, +3
- DMD, IVS57, G-C, -1
- DMD, LEU54ARG
- DMD, EX1DEL
- DMD, IVS26, T-G, +2
- DMD, GLN673TER
- DMD, 1-BP DEL, 10334C AND IVS69, G-T, +1
- DMD, IVS1, G-T, +1
- DMD, CYS3340TYR
- DMD, IVS2, G-T, -1
- DMD, 2-BP DEL, 382AG
- DMD, GLN60TER
- DMD, 1-BP INS, 402A
- DMD, GLN85TER
- DMD, ARG145TER
- DMD, ALA168ASP
- DMD, 1-BP DEL, 724C
- DMD, TYR231ASN
- DMD, GLN242TER
- DMD, GLU250TER
- DMD, 11-BP DEL, NT989
- DMD, 1-BP INS, NT1554
- DMD, GLY480TER
- DMD, GLN497TER
- DMD, TRP651TER
- DMD, LYS770TER
- DMD, LYS773GLU
- DMD, 52-BP DEL
- DMD, 1-BP INS, NT2928
- DMD, GLN1041TER
- DMD, TRP1063TER
- DMD, GLN1405TER

DMD, GLN1472TER

DMD, ARG1967TER

DMD, 1-BP DEL, 6408C

DMD, ARG2098TER

DMD, GLN2125TER

DMD, 17-BP DEL, NT6982

DMD, GLN2264TER

DMD, 1-BP INS, 7188A

DMD, IVS47, G-A, +1, EX48DEL

DMD, GLU2468TER

DMD, GLU2910VAL

DMD, ASN2912ASP

DMD, HIS2921ARG

DMD, SER3066TER

DMD, 4-BP DEL, NT9679

DMD, IVS65, G-A, +1

DMD, ARG3381TER

DMD, IVS70, G-A, +1

DMD, IVS70, G-T, +5

DMD, ALA3421VAL

DMD, 1-BP DEL, 10683C

DMD, 8-BP DEL, 1-BP INS, NT10692

DMD, THR279ALA

DMD, GLU1211TER, 3839G-T

DMD, ALU INS

DMD, ARG3190TER

DMD, ARG1314TER

DMD, 1-BP DEL, 377A

DMD, 16-BP DEL

DMD, IVS62, A-G, -285

DMD, IVS25, A-G, +2036

DMD, ARG2905TER

DMD, IVS2, T-A, +5591

DMD, TYR1995TER

DMD, EX45-47DEL

DMD, TRP3TER

FANCB, 1-BP INS, 1838T

FANCB, 3314-BP DEL

FANCB, 1-BP DEL, 1650T

FANCB, 1-BP INS, 811T

FANCB, IVS7DS, G-A, +5

FANCB, LEU717TER

FANCB, 2-BP DEL, 1857AG

FGD1, 1-BP INS, 2122G

FGD1, ARG610GLN

FGD1, ARG522HIS

FGD1, EX9-12DEL

FGD1, PRO312LEU

FGD1, 1-BP INS, 528C

FGD1, ARG408GLN

FGD1, 1-BP DEL, 2189A

FGD1, ARG433LEU

FGD1, 1-BP INS, 945C

FGD1, MET466VAL

FGD1, ARG656TER

FLNA, GLN182TER

FLNA, IVS4DS, T-C, +2

FLNA, IVS3AS, C-G, -3

FLNA, IVS2DS, G-A, +1

FLNA, 5-BP DEL, NT287

FLNA, LEU656PHE

FLNA, 5915C-G

FLNA, GLU82VAL

FLNA, PRO207LEU

FLNA, GLU254LYS

FLNA, ASP1159ALA

FLNA, ALA1188THR

FLNA, SER1199LEU

FLNA, 7315C-A

FLNA, SER1186LEU

FLNA, 9-BP DEL, NT4904

FLNA, 1-BP DEL, 2762G

FLNA, 1-BP DEL, 4147G

FLNA, ALA39GLY

FLNA, ASP203TYR

FLNA, ALA128VAL

FLNA, GLY1728CYS

FLNA, 1923C-T

FLNA, 2-BP DEL, 65AC

FLNA, ARG196TRP

FLNA, CYS210PHE

FLNA, PRO1291LEU

FLNA, 5217G-A, 48-BP DEL

FLNA, PRO637GLN

FLNA, GLY288ARG

FLNA, VAL711ASP

FLNA, 1,944-BP DEL

FLNA, TRP2632TER

FMR1, ILE304ASN

FMR1, 1-BP DEL, 373A

FMR1, IVS2AS1, G-T, -1 AND G-A, +1

FMR1, (CGG)n EXPANSION

FMR1, SER27TER

FTSJ1, EX9DEL

FTSJ1, 196C-T

FTSJ1, IVS2, G DEL, +1

FTSJ1, IVS3AS, A-G, -2

GDI1, LEU92PRO

GDI1, ARG70TER

GDI1, ARG423PRO

GDI1, 2-BP DEL, 1185AG

GK, IVS6AS, G-C, -1

GK, EX17DEL

GK, ASP440VAL

GK, 20-KB DEL

GK, ARG413TER

GK, TRP503ARG

GK, ALU INS, IVS4

GK, ASN288ASP

GPC3, 13-BP DEL, NT391

GPC3, EX7DEL

GPC3, TRP296ARG

GPC3, IVS5, G-T, +1

GPC3, ARG199TER

GPC3, HIS558TYR

GPC3, ALA1902THR

GPC3, EX6DEL

GPC3, IVS2, G-A, +1

GPC3, ARG387TER

GPC3, GLY556ARG

GRIA3, GLY833ARG

GRIA3, ARG631SER

GRIA3, MET706THR

GRIA3, 0.4-MB DEL

HCCS, 8.6-KB DEL

HCCS, ARG197TER

HCCS, ARG217CYS

HCFC1, 455A-G

HCFC1, SER225ASN

HDAC8, IVS1DS, 164+5G-A

HDAC8, ARG164TER

HDAC8, HIS180ARG

HDAC8, THR311MET

HDAC8, GLY320ARG

HDAC8, HIS334ARG HPRT, ILE132MET

HPRT, ASP80VAL

III KI, ASI 60 VAL

HPRT, ASP201GLY

HPRT, 1-BP INS, 56T

HPRT, EX8DEL

HPRT, LEU41PRO

HPRT, 24AA+

HPRT, PHE74LEU

HPRT, ASP194ASN AND ASP193ASN

HPRT, SER110LEU

HPRT, 3-BP DEL, VAL179DEL

HPRT, VAL130ASP

HPRT, ALA161SER

HPRT, SER104ARG

HPRT, PHE199VAL

HPRT, GLY70GLU

HPRT, GLY71ARG

HPRT, GLN108TER

HPRT, HIS203ASP

HPRT, ARG44LYS

HPRT, ASP176TYR

HPRT, 2-BP DEL, GT

HPRT, 1-BP DEL, TTA-TA

HPRT, 1-BP DEL, TTG-TG

HPRT, 40-BP DEL

HPRT, IVS8DS, G-A, +5

HPRT, IVS8AS, ATAG-TTTG

HPRT, IVS7DS, G-A, +5

HPRT, IVS1AS, A-T, -2

HPRT, PRO176LEU

HPRT, ARG51GLY

HPRT, ARG51TER

HPRT, MET56THR

HPRT, MET143LYS

HPRT, ARG170TER

HPRT, 13-BP DEL, 5-PRIME UTR

HPRT, EX2DEL

HPRT, EX4-9DEL

HPRT, EX6-9DEL

HPRT, EX9DEL

HPRT, DEL

HPRT, INV/DEL, EX6-9

HPRT, EX2-3DUP, IVS1DEL

HPRT, THR168ILE

HPRT, GLY16SER

HPRT, GLY58ARG

HPRT, LEU78VAL

HPRT, EX6DEL

HPRT, 1-BP INS, 14823T

HPRT, ASP52GLY

HPRT, GLY140ASP

HPRT, ASP194GLU

HPRT, TYR153TER

HPRT, 2969-BP DEL, NT970

HPRT, LEU65PHE

HPRT, ARG48HIS

HSD17B10, ARG130CYS

HSD17B10, LEU122VAL

HSD17B10, ASN247SER

HSD17B10, ARG192ARG

HSD17B10, GLU249GLN

HUWE1, ARG4013TRP

HUWE1, ARG2981HIS

HUWE1, ARG4187CYS

*HUWE1, VAL950ASP

*HUWE1, DUP

IDS, ARG443TER

IDS, SER333LEU

IDS, TRP502SER

IDS, PRO160ARG

IDS, ARG172TER

IDS, 60-BP DEL, NT1244

IDS, DEL

IDS, CYS422GLY

IDS, LYS135ARG

IDS, TRP475TER

IDS, 2-BP DEL, CODON 170

IDS, ARG468TRP

IDS, ARG468GLN

IDS, 78-BP INS

IDS, ARG468LEU

IDS, 3-BP DEL, 473TCC

IDS, GLY489ALA, MET488ILE

IGBP1, -57delT and -55T-A, 5-PRIME UTR

IKBKG, EXON 4-10 DEL

IKBKG, TER420TRP

IKBKG, 10-BP INS, NT127

IKBKG, 1-BP INS, 1110C

IKBKG, MET407VAL

IKBKG, PRO62TER

IKBKG, GLU391TER

IKBKG, 1-BP DUP, 1167C

IKBKG, CYS417ARG

IKBKG, CYS417PHE

IKBKG, ASP406VAL

IKBKG, 13-BP DUP, NT1166

IKBKG, 4.4-KB DUP

IKBKG, LEU153ARG

IKBKG, GLN403TER

IKBKG, IVS6DS, G-A, +5

IKBKG, 1-BP INS, 1409A

IKBKG, IVS8, -1, G-A

IKBKG, 1-BP INS, 110C

IKBKG, ALA288GLY

IKBKG, GLU315ALA

IKBKG, ARG319GLN

IKBKG, 518C-G, ARG173GLY

IL1RAPL1, TYR459TER

IL1RAPL1, TRP487TER

IL1RAPL1, EX2-5DEL

IL1RAPL1, 730-KB DEL, EX3-7

IQSEC2, ARG863TRP

IQSEC2, GLN801PRO

IQSEC2, ARG758GLN

IQSEC2, ARG359CYS

IQSEC2, ARG855TER

KDM5C, LEU731PHE

KDM5C, 1-BP INS, 202C

KDM5C, ALA388PRO

KDM5C, ARG694TER

KDM5C, SER451ARG

KDM5C, ARG766TRP

KDM5C, ALA77THR

KDM5C, CYS724TER

KDM5C, PRO554THR

SHROOM4, SER1089LEU

*SHROOM4, GLU474GLU

L1CAM, IVS18AS, A-C, -19

L1CAM, CYS264TYR

L1CAM, 1.3-KB DUP

L1CAM, HIS210GLN

L1CAM, ASP598ASN

L1CAM, GLY452ARG

L1CAM, ARG184GLN

L1CAM, 2-BP DEL, EX26

L1CAM, SER1194LEU

L1CAM, ILE179SER

L1CAM, GLY370ARG

L1CAM, 2-BP DEL, EX18

L1CAM, 924C-T

L1CAM, VAL752MET

L1CAM, IVS15DS, G-A, +5

L1CAM, GLN974TER

L1CAM, PRO240LEU

L1CAM, IVS26AS, G-C, -1

LAMP2, 2-BP DEL, 1097AA

LAMP2, LEU113TER

LAMP2, IVS6, G-C, +5

LAMP2, 1-BP INS, 974A

LAMP2, IVS5, G-A, +1

LAMP2, 1-BP DEL, 14G

LAMP2, 1-BP INS, 883T

LAMP2, 7-BP DEL

LAMP2, GLN174TER

LAMP2, VAL310ILE

LAMP2, TRP321ARG

LAMP2, 1-BP DEL, 1219A

MAGT1, VAL343GLY

MAOA, GLN296TER

MAOA, PROMOTER POLYMORPHISM, 30-BPREPEAT

MBTPS2, HIS277LEU

MBTPS2, MET87ILE

MBTPS2, ARG429HIS

MBTPS2, PHE475SER

MBTPS2, TRP226LEU

MBTPS2, ARG508SER

MECP2, ARG133CYS

MECP2, PHE155SER

MECP2, 1-BP DEL, 806G

MECP2, 44-BP DEL, NT1152

MECP2, ARG270TER

MECP2, IVS2, A-G, -2

MECP2, THR158MET

MECP2, ARG106TRP

MECP2, GLU406TER

MECP2, 2-BP DEL, 211CC

MECP2, ARG294TER

MECP2, 41-BP DEL, NT1157

MECP2, 41-BP DEL, NT1159

MECP2, 44-BP DEL, NT1159

MECP2, ALA140VAL

MECP2, ARG306CYS

MECP2, GLU137GLY

MECP2, 1-BP DEL, 76C

MECP2, 14-BP DUP, NT766

MECP2, ARG168TER

MECP2, ARG255TER

MECP2, 240-BP DEL, NT1161

MECP2, GLY428SER

MECP2, 52-BP DEL

MECP2, TYR141TER

MECP2, GLU455TER

MECP2, LEU100VAL

MECP2, 11-BP DEL, EX1

MECP2, 5-BP DUP, EX1

MECP2, DUP

MECP2, 1-BP DEL AND 2-BP INS

MECP2, 2-BP DEL, 488GG

MECP2, PRO225LEU

MECP2, 32-BP DEL, NT1154

MECP2, PRO322SER

MECP2, PRO152ALA

MECP2, ALA2VAL

MECP2, 1-BP DEL, 710G

MED12, ARG961TRP

MED12, ASN1007SER

*MED12, ARG1148HIS

*MED12, SER1165PRO

*MED12, HIS1729ASN

MID1, 3-BP DEL, MET438

MID1, 24-BP DUP

MID1, 1-BP INS

MID1, LEU626PRO

MID1, GLU115TER

MID1, EX1 DUP

MID1, LEU295PRO

MID1, 2-BP DEL, 1545GA

MID1, GLU238TER

NAA10, SER37PRO

*NAA10, ARG116TRP

NDP, ARG90PRO

NDP, SER75CYS

NDP, VAL60GLU

NDP, TYR44CYS

NDP, CYS96TYR

NDP, LEU124PHE

NDP, CYS69SER

NDP, CYS128TER

NDP, MET1VAL

NDP, ARG121TRP

NDP, LEU13ARG

NDP, LEU61PHE

NDP, HIS42ARG

NDP, 1-BP DEL

NDP, ALA105THR

NDP, CYS110GLY

NDP, ARG121LEU

NDP, CYS96TRP

NDP, VAL45GLU

NDP, SER73TER

NDP, SER101PHE

NDUFA1, GLY8ARG

NDUFA1, ARG37SER

NDUFA1, GLY32ARG

NHS, 1-BP INS, 2387C

NHS, 1-BP DEL, 3459C

NHS, ARG378TER

NHS, 1-BP INS, 718G

NHS, GLN39TER

NHS, IVS3AS, A-G, -2

NHS, 500-KB TRIPLICATION

NHS, 4.8-KB DEL

NLGN3, ARG451CYS

NLGN4, 1-BP INS, 1186T

NLGN4, 2-BP DEL, 1253AG

NLGN4, 757-KB DEL

NSDHL, ALA105VAL

NSDHL, GLY205SER

NSDHL, GLN210TER

NSDHL, ARG88TER

NSDHL, ALA182PRO

NSDHL, GLU151TER

NSDHL, 1-BP DUP, 1098T

NSDHL, 3-BP DEL, 696GAA

OCRL, 112-BP DEL

OCRL, ARG-TER

OCRL, ARG577GLN

OCRL, HIS601GLN

OCRL, TYR462CYS

OCRL, ARG301CYS

OCRL, ARG476TRP

OCRL, ILE526THR

OCRL, 2-BP DEL, 166TT

OFD1, SER434ARG

OFD1, 1-BP DEL, 312G

OFD1, 19-BP DEL, NT294

OFD1, IVS5AS, T-G, -10

OFD1, 2-BP INS, 1887AT

OFD1, 4,094-BP DEL, 14-BP DEL

OFD1, 4-BP DUP, 2122AAGA

OFD1, 7-BP DEL, NT2841

OFD1, 1-BP DEL, 2767G

OFD1, 18-BP DEL, EX 8

OPHN1, 1-BP DEL, NT1578

OPHN1, 8-BP DUP

OPHN1, GLN62TER

OPHN1, 17.6-KB DEL

OPHN1, 2-BP DEL, NT642

OPHN1, 68-KB DEL

OTC, DEL

OTC, ARG109GLN

OTC, ARG109TER

OTC, LEU111PRO

OTC, GLN216GLU

OTC, GLU154TER

OTC, LEU45PRO

OTC, ARG26GLN

OTC, LYS46ARG

OTC, ARG245TRP

OTC, GT-GC, INTRON 7

OTC, GTA-GTG, INTRON 7

OTC, IVS4, A-T, -2

OTC, ARG277TRP

OTC, PRO225LEU

OTC, GLU87LYS

OTC, GLY50TER

OTC, GLY162ARG

OTC, 1-BP DEL, 403G

OTC, IVS2, G-A, -1

OTC, GLY47GLU

OTC, ARG62THR

OTC, LEU272PHE

OTC, TYR313ASP

OTC, ARG129HIS

OTC, LEU148PHE

OTC, MET206ARG

OTC, ARG40CYS

OTC, ARG40HIS

PAK3, ARG419TER

PAK3, ARG67CYS

PAK3, ALA365GLU

PAK3, TRP446SER

PAK3, IVS6DS, A-G, +4

PCDH19, 1-BP INS, 1091C

PCDH19, VAL441GLU

PCDH19, GLN85TER

PCDH19, SER671TER

PCDH19, 1-BP INS, 2030T

PCDH19, GLU48TER

PCDH19, 5-BP DUP, NT1036

PCDH19, ASN557LYS

PDHA1, 4-BP DEL

PDHA1, 7-BP DEL

PDHA1, ARG378HIS

PDHA1, LYS313DEL

PDHA1, 2-BP DEL

PDHA1, 20-BP DEL, EX11DEL

PDHA1, 21-BP INS

PDHA1, ARG234GLY

PDHA1, ARG302CYS

PDHA1, 4-BP INS, 1251ACTA

PDHA1, ASP258ALA

PDHA1, PHE205LEU

PDHA1, TYR243ASN

PDHA1, ASP315ASN

PDHA1, MET282LEU

PDHA1, 1-BP INS, FS141TER

PDHA1, ARG10PRO

PDHA1, 13-BP INS, EX10

PDHA1, 36-BP INS

PDHA1, ARG288HIS

PDHA1, 12-BP INS, EX11

PDHA1, ARG263GLY

PDHA1, LEU216PHE

PGK1, ASP268ASN

PGK1, ARG206PRO

PGK1, VAL266MET

PGK1, THR352ASN

PGK1, LEU88PRO

PGK1, GLY157VAL

PGK1, CYS315ARG

PGK1, 3-BP DEL, LYS191DEL

PGK1, ILE252THR

PGK1, ASP285VAL

PGK1, ILE46ASN

PGK1, SER319ASN

PGK1, ASP164VAL

PGK1, IVS7DS, G-A, +5

PGK1, THR378PRO

PHF6, ARG342TER

PHF6, CYS99PHE

PHF6, LYS234GLU

PHF6, CYS45TYR

PHF6, HIS229ARG

PHF6, MET1TYR

PHF6, ARG257GLY

PHF6, LYS8TER

PHF6, IVS2AS, A-G, -8

PHF6, 1-BP INS, 27A

PHF8, 12-BP DEL

PHF8, ARG211TER

PHF8, LYS177TER

PHF8, PHE279SER

PLP1, PRO215SER

PLP1, TRP162ARG

PLP1, PRO14LEU

PLP1, THR155ILE

PLP1, VAL218PHE

PLP1, DEL

PLP1, THR181PRO

PLP1, LEU223PRO

PLP1, ASP202HIS

PLP1, GLY73ARG

PLP1, GLY220CYS

PLP1, HIS139TYR

PLP1, ILE186THR

PLP1, THR42ILE

PLP1, MET1ILE

PLP1, -34C-T, 5-PRIME UTR

PLP1, PHE236SER

PLP1, TRP144TER

PLP1, ALA242VAL

PLP1, SER169PHE

PLP1, DUP

PLP1, IVS6DS, G-T, +3

PLP1, IVS3DS, T-C, +2

PLP1, IVS3DS, A-G, +4

PLP1, IVS3, 19-BP DEL, +28

PLP1, ARG137TRP

PLP1, ASP57TYR

PORCN, 13-BP DUP, NT1059

PORCN, 178G-A, GLY60ARG

PORCN, ARG124TER

PORCN, TRP74TER

PORCN, ARG365GLN

PQBP1, 2-BP INS, 3900AG

PQBP1, 4-BP DEL, 3896AGAG

PQBP1, 2-BP DEL, 3898AG

PQBP1, 1-BP INS, 641C

PQBP1, 23-BP DEL, NT547

PQBP1, 21-BP DEL, NT334

PQBP1, TYR65CYS

PRPS1, ASN113SER

PRPS1, ASP182HIS

PRPS1, ASP51HIS

PRPS1, LEU128ILE

PRPS1, ALA189VAL

PRPS1, HIS192GLN

PRPS1, GLU43ASP

PRPS1, MET115THR

PRPS1, LEU152PRO

PRPS1, GLN133PRO

PRPS1, ASP65ASN

PRPS1, ALA87THR

PRPS1, GLY306ARG

PRPS1, ILE290THR

PRPS1, VAL142LEU

*PTCHD1, I173V

*PTCHD1, ML336-7II

*PTCHD1, E479G

*PTCHD1, L73F

*PTCHD1, A470D

*PTCHD1, H359R

*PTCHD1, V195I

RAB39B, IVS1DS, G-A, +1

RAB39B, TYR7TER

RBM10, 1-BP INS, 1893A

RBM10, TRP412TER

RPL10, LEU206MET

RPL10, HIS213GLN

RPS6KA3, 187-BP DEL, NT406

RPS6KA3, GLY75VAL

RPS6KA3, SER227ALA

RPS6KA3, VAL82PHE

RPS6KA3, IVS4AS, G-C, -1

RPS6KA3, ARG114TRP

RPS6KA3, 2-BP DEL, 451AG

RPS6KA3, GLN689TER

RPS6KA3, ARG729GLN

RPS6KA3, ARG383TRP

RPS6KA3, ILE189LYS

RPS6KA3, IVS6, A-G, +3

RPS6KA3, IVS5, A-G, -11

RPS6KA3, 1-BP DEL, 2144C

RPS6KA3, IVS12, A-G, -2

RPS6KA3, IVS3, L1 INS, -8

RPS6KA3, PHE268SER

RPS6KA3, 3-BP DEL, 1428TAT

RPS6KA3, DUP EXONS 17-20, NT1959

RPS6KA3, 3-BP DEL, 454GGA

RPS6KA3, THR115SER

*SIZN1, ARG7CYS

*SIZN1, THR344ILE

SLC16A2, LEU512PRO

SLC16A2, 1-BP DEL, 1212T

SLC16A2, ALA224VAL

SLC16A2, EX1DEL

SLC16A2, LEU397PRO

SLC16A2, 2.4-KB DEL

SLC16A2, LEU568PRO

SLC16A2, LEU434TRP

SLC16A2, SER448TER

SLC16A2, PHE230 DEL

SLC16A2, 1-BP DEL, 1834C

SLC6A8, ARG514TER

SLC6A8, GLY381ARG

SLC6A8, 3-BP DEL, 1221TTC

SLC6A8, 1-BP INS, 950A

SLC6A8, GLY87ARG

SLC6A8, IVS1AS, A-G, -2

SLC6A8, CYS337TRP

SLC6A8, GLY132VAL

SLC6A8, CYS491TRP

SLC6A8, 3-BP DEL, 1006AAC

SLC9A6, 6-BP DEL, NT764

SLC9A6, ARG468TER

SLC9A6, IVS3, AA-CC

SLC9A6, 2-BP DEL, 511_512delAT

SLC9A6, 9-BP DEL, NT1012

SMC1A, 3-BP DEL, 2493CCA

SMC1A, GLU493ALA

SMC1A, 15-BP DEL, NT173

SMC1A, ARG496HIS

SMC1A, ILE784THR

SMC1A, 8.152-KB DEL

SMS, IVS4AS, G-A, +5

SMS, GLY56SER

SMS, VAL132GLY

SOX3, 33-BP DUP, NT711-743

SOX3, DUP

SOX3, 21-BP DUP

SRPX2, ASN327SER

SRPX2, TYR72SER

SYN1, TRP356TER

*SYN1, GLN555TER

*SYN1, ALA51GLY

*SYN1, ALA550THR

*SYN1, THR567ALA

SYP, 1-BP INS, 274A

SYP, 2-BP DEL/INS

SYP, 4-BP DEL, 829GACT

SYP, GLY217ARG

TIMM8A, 1-BP DEL, 151T

TIMM8A, 10-BP DEL, FS61TER

TIMM8A, GLU24TER

TIMM8A, CYS66TRP

TIMM8A, 1-BP DEL, 108G

TIMM8A, DEL

TIMM8A, ARG80TER

TIMM8A, IVS1, A-C, -23

TIMM8A, 1-BP DEL, 127T

TSPAN7, GLY218TER

TSPAN7, PRO172HIS

TSPAN7, 2-BP DEL, 564GT

UBE2A, GLN128TER

UBE2A, GLY23ARG

UBE2A, ARG11GLN

*UBE2A, PHE72SER

UPF3B, 4-BP DEL, 674GAAA

UPF3B, 2-BP DEL, 867AG

UPF3B, ARG430TER

UPF3B, TYR160ASP

ZDHHC9, 4-BP DUP, 172CGCT

ZDHHC9, 167+5G-C

ZDHHC9, ARG148TRP

ZDHHC9, PRO150SER

ZNF41, PRO111LEU

ZNF41, IVS2AS, A-C, -42

ZNF674, GLU118TER

*ZNF674, PRO412LEU

*ZNF674, MET343THR

ZNF711, 2-BP DEL, 2157TG ZNF711, ARG525TER ZNF81, SER179ASN

Table S3. List of All XLID Mutations and Variants Affecting the Same Amino Acid as a Reported XLID Mutation, which Are Listed in EVS

Chr position	Gene	mRNA Accession#	XLID-mutatio Variant found in		AA Position/ Length total	Pred	yphen diction 1 vs EVS)	Grant Scor (OMI EVS	re [M/	MAF (% EA/ AA/ All)	All Genotypes	Cons. (Phast Cons)	Cons. (GERP)	Ref & comments
a) XLID	mutations pre	esent in EVS												
X:115303595	AGTR2 ^a	NM_000686.4	c.62G>T p.Gly21Val (rs121917810)	21/364	benign	109	0.46(0.0522/ ()/ TG=23 -4037/ G		0.0	3.6	590 ID males were screened after the identification of <i>AGTR2</i> as an ID candidate due to its disruption in a translocation in an individual. p.G21V was present in the proband and his affected brother and in a third unrelated individual. F4 57 males with NS-ID were screened 1 was carrying p.G21V (but no segregation study was done) F5 Reported in 4 individuals when screening 908 hemochromatosis individuals, suggesting it is a polymorphism. F6
X:115304504	AGTR2 ^a	NM_000686.4	c.971G>A p.Arg324Gln (<i>rs35474657</i>)	324/364	benign	43	0.0/ 0.7 0.26			0/ AG=2- =4036/ G		1.0	1.7	590 ID males were screened after the identification of <i>AGTR2</i> as an ID candidate due to its disruption in a translocation in a male. p.R324Q was detected in 3 unrelated cases and was not found in 510 control X-chromosomes. ⁵⁴
X:135861667	ARHGEF6 ^a	NM_004840.2	c.166-11T>C p.? (rs140322310)	-	-	-	0.237 0.0522/ (0/ GA=1: -4047/ A		0.16	4.48	Screening of 119 additional individuals with NS-ID after its identification as an ID candidate led to the identification of this variation. It was reported in all affected males from the family and 5 carrier unaffected females. The predicted effect on splice efficiency is marginal (79% versus 77%), but results in exon 2 skipping. It is absent from 170 control X-chromosomes. 57
X:41448815	CASK	NM_003688.3	c.1186C>T p.Pro396Ser (rs137852820)	396/922	benign	74	0.0/ 0.0			A=0/ AC =4057/ G		1.0	5.8	Reported in 3 individuals: 2 affected brothers and 1 affected female maternal cousin. It was not detected in 390 control X-chromosomes. The cosegregation study was not done in the healthy sister, who had 2 affected sons. Associated LOD score: 0.12. ²
X:31496431	DMD	NM_004006.2	c.8729A>T p.Glu2910Val (rs41305353)	2910/3686	benign	121	0.847 6.8615/			/ AT=249 =3803/ T		1.0	5.4	One proband presenting with intermediate DMD phenotype was reported to carry two variations (p.E2910V, p.N2912D). The sister carried all 3 variants (p.E2910V, p.N2912D, p.H2921R). The mother carried only p.H2921R, and the father none. Germinal mosaicism? Already supposing that at least 1 variant is not fully pathogenic. ⁵⁸

X:31496426	DMD	NM_004006.2	c.8734A>G p.Asn2912Asp (rs1800278)	2912/3686	benign	23	0.8472/ 7.2006/ 3.1531	CC=9/ CT=255/ C=60/ TT=3795/ T=2383	1.0	1.7	One male presenting with intermediate DMD phenotype was reported to carry two variations (p.E2910V, p.N2912D). The sister carried all 3 variants (p.E2910V, p.N2912D, p.H2921R). The mother carried only p.H2921R, and the father none. Germinal mosaicism? Already supposing that at least 1 variant is not fully pathogenic. ⁵⁸
X:31496398	DMD	NM_004006.2	c.8762A>G p.His2921Arg (rs1800279)	2921/3686	benign	29	2.9727/ 0.4435/ 2.0547	CC=2/ CT=164/ C=49/ TT=3893/ T=2394	1.0	2.9	One male presenting with intermediate DMD phenotype was reported to carry two variations (p.E2910V, p.N2912D). The sister carried all 3 variants (p.E2910V, p.N2912D, p.H2921R). The mother carried only p.H2921R, and the father none. Germinal mosaicism? Already supposing that at least 1 variant is not fully pathogenic. ⁵⁸
X:54496615	FGD1	NM_004463.2	c.935C>T p.Pro312Leu (rs28935498)	312/962	unknown	98	0.1041/ 0.0/ 0.0663	AA=0/ AG=5/ A=2/ GG=4054/ G=2438	1.0	3.8	Three brothers carried the mutation, maternally transmitted. The mother was the only sample available for co-segregation testing. The observed phenotype was not suggestive of classical Aarskog syndrome. However, it was predicted to eliminate a β-turn, which may affect the orientations of an SH3 binding domain. ⁵⁹
X:153588207	FLNA	NM_001456.3	c.3872C>T p.Pro1291Leu (rs137853319)	1291/2640	benign	98	0.0153/ 0.0/ 0.0099	AA=0/ AG=0/ <mark>A=1/</mark> GG=3859/ G=2375	0.9	5.3	The variant was transmitted by the affected mother and absent from 100 control X-chromosomes. It did not affect a very conserved residue, but the phenotype of the proband was somewhat similar to the one described by Hehr et al. (6) (periventricular nodular heterotopia, craniofacial dysmorphic features and severe constipation).
X:70444261	GJB1	NM_000166.5	c.704T>G p.Phe235Cys (rs104894825)	235/284	benign	205	0.0/ 0.8344/ 0.3031	GG=0/ GT=28/ G=4/ TT=4030/ T=2439	0.854	4.9	This variant was identified in a girl with unusually more severe neuropathy matching rather Dejerine—Sottas disease than classical CMTX. It was inherited from her unaffected mother who showed unusual X-inactivation pattern. It was absent from 50 control chromosomes and detected in 6 additional CMTX samples (out of 12,720). The localization and trafficking of the mutant protein in cell culture was normal, but electrophysiological studies showed that the mutation caused abnormal hemichannel opening with excessive permeability of the plasma membrane and decreased cell survival. 62
X:77086362	MAGT1 ^a	NM_032121.5	c.1028T>G p.Val343Gly/p.Val311Gly (rs145245774)	343/368	possibly-damaging	109	0.1785/ 0.0261/ 0.1231	CC=0/ CA=8/ C=5/ AA=4051/ A=2434	0.989	5.49	This missense variant was identified in one male and his affected brother and nephew, and not in the non-affected sister. The mutation was annotated p.G311V but corresponds in fact to the p.G343V described in EVS (see electropherogram profile in the paper). This variant was not found in 267 control X-chromosomes. 63

X:119005968	NDUFA1	NM_004541.3	c.94G>C p.Gly32Arg (rs1801316)	32/71	Probably-damaging	125	0.9958/ 0.1043/ 0.6722	CC=0/ CG=53/ C=18/ GG=4007/ G=2425	0.8	3.8	It was identified in two independent families. Complex I was specifically reduced to 5% to 10% residual activity in both unrelated probands. It was inherited from both carrier mothers, and absent from 150 controls even though it was already reported in dbSNP. ⁶⁴
X:1937 <i>5</i> 782	PDHA1	NM_000284.3	c.844A>C p.Met282Leu (rs2229137)	282/391	benign	15	0.0595/ 0.2086/ 0.1136	CC=0/ CA=8/ C=4/ AA=4052/ A=2439	1.0	5.5	This missense variant was found in 1 female (dysmorphy, microcephaly, hypotonia, brain atrophy) with PDH activity within normal range, and X inactivation (80:20). She was born from consanguineous parents. She presented with elevated lactate, but normal amount of immunoreactive protein. Her brother showed a more severe phenotype but DNA sample was unavailable because of his early death. ⁶⁵
X:23397873	PTCHD1 ^a	NM_173495.2	c.517A>G p.Ile173Val (<i>rs147324438</i>)	173/889	benign	29	0.0595/ 0.0/ 0.0379	GG=0/ GA=2/ G=2/ AA=4058/ A=2441	1.0	5.06	This missense variant was reported in a male with autism, inherited from his unaffected mother. It was absent from 1'100 control individuals. The proband did not have siblings, which did not allow to support or exclude the pathogenicity of this variant. ⁴⁴
X:23353209	PTCHD1 ^a	NM_173495.2	c.217C>T p.Leu73Phe	73/889	probably-damaging	22	0.0/ 0.0261/ 0.0095	TT=0/ TC=1/ CC=4059/ C=2443	1.0	4.46	This variant was reported in a male with autism, inherited from his unaffected mother. It was absent from 869 control individuals. This variant did not segregate with the ASD status in the family since it was present in a supposedly unaffected brother, and absent from an affected one, already suggesting it might not be the causative mutation. ⁴⁴
X:99922289	SRPX2 ^a	NM_014467.2	c.980A>G p.Asn327Ser (rs121918363)	327/466	benign	46	0.1635/ 0.0/ 0.1041	GG=0/ GA=8/ G=3/ AA=4052/ A=2440	1.0	5.5	This variant was the first mutation identified in <i>SRPX2</i> and was absent from 554 control X-chromosomes. It co-segregated with the disease phenotype since it was present in all affected females and transmitted by the affected maternal grandfather. It was shown to affect the protein N-glycosylation. ⁶⁶
X:99917224	SRPX2 ^a	NM_014467.2	c.215A>C p.Tyr72Ser (rs121918364)	72/466	benign	144	0.0149/ 0.0/ 0.0095	CC=0/ CA=0/ C=1/ AA=4060/ A=2442	1.0	5.1	This variant was the second mutation identified in <i>SRPX2</i> when screening 172 additional subjects. It was transmitted by the healthy mother, present in 2 affected aunts and also in an unaffected aunt. It was absent from 624 controls. ⁶⁶
X:47478976	SYN1 ^a	NM_006950.3	c.152C>G p.Ala51Gly	51/706	possibly-damaging	60	0.0219/ 1.7629/ 0.6007	CC=0/ CG=30/ C=11/ GG=2659/ G=1436	0.718	3.08	This missense variant was detected in one epileptic female without cognitive impairment and in one ASD male also carrying the p.T567A variant, already suggesting one of the two variants might not contribute to the phenotype. 48
X:38535032	TSPAN7	NM_004615.3	c.515C>A p.Pro172His (rs104894951)	172/250	benign	77	0.1338/ 0.0/ 0.0852	AA=0/ AC=8/ A=1/ CC=4051/ C=2442	0.6	3.6	This variant was identified in one XLID family and cosegregated with the disease in all but 1 affected individual. After neurological reassessment, linkage analysis excluded this gene ⁶⁷ It was then confirmed that the linkage data did not match with a mutation in <i>TSPAN7</i> in 105 ID

											males revealed again the p.P172H variant. The mother and the sister were both unaffected carriers. ⁶⁹
X:47308837	ZNF41 ^a	NM_007130.2	c.332C>T p.Pro111Leu (rs104894955)	111/780	benign	98	0.1486/ 0.0/ 0.0947	AA=0/ AG=8/ A=2/ GG=4051/ G=2441	1.0	3.1	The screening of <i>ZNF41</i> in 210 XLID cases revealed this missense variant in one proband. It was also present in one affected brother, transmitted byt the mother. No additional samples were available to do a full cosegregation analysis. It was not found in 401 controls. ⁷⁰
X:47315839	ZNF41 ^a	NM_153380.2	c.73-42 p.?	-	-	-	0.0297/ 0.3129/ 0.1325	GG=0/ GT=9/ G=5/ TT=4051	0.197	-1.56	The screening of <i>ZNF41</i> in 210 XLID cases revealed this variant in one proband. It was transmitted by the mother, and also present in a mildly affected sister. However, no difference of expression could be detected in the predominant <i>ZNF41.1</i> transcript, just an additional transcript was present. It was not found in 405 controls. ⁷⁰
X:117959226	ZCCHC12 a	NM_173798.2	c.19C>T p.Arg7Cys (rs35356061)	4/403	benign	180	0.0149/ 0.8344/ 0.3124	TT=0/ TC=28/ T=5/ CC=4032/ C=2438	0.0080	2.17	This variant was identified during the screening of a cohort of 729 ID males and 494 controls. It was detected in 4/729 males with ID and 1/494 controls. In one family, this variant did not segregate with the disease status, and in the 3 other families no DNA sample was available from the parents. The observed residual function of the protein was showing 40% of activity. All those information already suggested it was a polymorphism. ⁶

b) XLID mutation and EVS variant affecting the same amino acid position

Chr position	Gene	mRNA Accession#	XLID-mutation	Variant found in EVS	AA Position/ Length total	Polyphen Prediction (OMIM vs EVS)	Grantham Score (OMIM/ EVS)	MAF (% EA/ AA/ All)	All Genotypes #	Cons. (Phast Cons)	Cons. (GERP)
X:153001649	ABCD1	NM_000033.3	c.1165C>G p.Arg389Gly (rs128624215)	c.1165C>T p.Arg389Cys	389/746	Probably- damaging/Probably- damaging	125/180	0.0/ 0.0261/ 0.0095	TT=0/ TC=1/ CC=4059/ C=2443	1.0	4.8

X:115303594	AGTR2 ^a	NM_000686.4	c.62G>T p.Gly21Val (rs121917810)	c.61G>A p.Gly21Arg	21/364	benign/benign	109/125	0.0149/ 0.0/ 0.0095	AA=0/ AG=1/ GG=4059/ G=2443	0.0	1.8
X:32716111	DMD	NM_004006.2	c.835A>G p.Thr279Ala (rs128627255)	c.836C>T p.Thr279Met	279/3686	Probably- damaging/Probably- damaging	58/81	0.0149/ 0.0/ 0.0095	AA=0/ AG=1/ GG=4057/ G=2437	1.0	5.6
X:21863324	MBTPS2	NM_015884.3	c.261G>A p.Met87Ile (rs122468177)	c.260T>C p.Met87Thr	87/520	Probably- damaging/Probably- damaging	10/81	0.0149/ 0.0/ 0.0095	CC=0/ CT=0/ C=1/ TT=4060/ T=2442	1.0	5.81
X:38240681	ОТС	NM_000531.5	c.386G>A p.Arg129His (rs66656800)	c.385C>T p.Arg129Cys (rs140046498)	129/355	Probably- damaging/Probably- damaging	29/180	0.0149/ 0.0261/ 0.0189	TT=0/ TC=2/ CC=4057/ C=2443	1.0	5.0
X:99922290	SRPX2 ^a	NM_014467.2	c.980A>G p.Asn327Ser (rs121918363)	c.981C>G p.Asn327Lys	327/466	benign/benign	46/94	0.0/ 0.0261/ 0.0095	GG=0/ GC=1/ CC=4059/ C=2443	1.0	-0.2
X:133609228	HPRT	NM_000194.2	c.151C>G p.Arg51Gly (rs137852494)	c.152G>A p.Arg51Gln	51/219	benign/benign	125/43	0.0/ 0.0261/ 0.0095	AA=0/ AG=0/ A=1/ GG=4060/ G=2442	1.0	4.7
X:99662275	PCDH19	NM_001184880.1	c.1322T>A p.Val441Glu (rs132630323)	c.1321G>C p.Val441Leu (rs200126728)	441/1149	Probably- damaging/Probably- damaging	121/32	0.0604/ 0.0/ 0.0388	GG=0/ GC=4/ CC=3955/ C=2395	0.9	5.9
X:73740842	SLC16A2	NM_006517.3	c.671C>T p.Ala224Val (rs104894936)	c.670G>A p.Ala224Thr	224/614	unknown/unknown	64/58	0.0149/ 0.0/ 0.0095	AA=0/ AG=0/ A=1/ GG=4060/ G=2442	0.5	4.7

^a: Genes further discussed in the paper. In red: variations reported in hemizygous males or homozygous females from the NHLBI cohort.

Table S4. Reported Frameshift Variants in EVS in Our Set of 104 Selected Genes, Supposedly Implicated in XLID When Mutated

Chr position	Gene	mRNA Accession #	Resulting frameshift	variation/ Reference	MAF (% EA/ AA/ All)	All Genotypes	HW	Read depth	Cons. (GERP)	Location (exon affected/# total exons) & Comments
X:5811528	NLGN4X	NM_020742.2	p.Leu593Phefs*7	A/ACGAG	0.36/ 1.2998/ 0.6997	A1A1=20/ A1R=1/ A1=29/ RR=3900/ R=2134	No	169	4.1	exon 7/7
X:13764945	OFD1	NM_003611.2	p.Lys237Serfs*6	C/CA	1.5743/ 0.9401/ 1.3429	A1A1=12/ A1R=48/ A1=65/ RR=3888/ R=2241	No	47	4.2	exon 8/23. del of a A in a stretch of As
X:99662504	PCDH19	NM_001105243.1	p.Tyr366Leufs*10	CG/C	0.1092/ 0.3808/ 0.2082	A1A1=0/ A1R=10/ A1=11 / RR=3899/ R=2259	No	47	-0.2	exon 1/6 . Insertion of a C in a stretch of 6Cs
X:54069205	PHF8 b	NM_001184896.1	p.Thr22Argfs*32	GTC/G	0.0/ 0.027/ 0.0098	A1A1=0/ A1R=0/ A1=1/ RR=3941/ R=2285	Yes	23	1.1	Alternative exon 2/22, in 5'UTR of other isoforms
X:48759667	PQBP1 ^b	NM_001032381.1	p.Glu154Alafs*12	C/CAG	1.7441/ 1.6931/ 1.7255	A1A1=14/ A1R=58/ A1=90/ RR=3877/ R=2212	No	44	-6.6	Alternative exon 5A. Del AG in a stretch of 6 AGs.
X:47478983	SYNI ^a	NM_006950.3	p.Ala49Profs*95	CG/C	0.4852/ 0.7704/ 0.5799	A1A1=9/ A1R=14/ A1=2/ RR=2321/ R=1173	No	3	2.0	exon 1/13

X:47306866	ZNF41 ^a	NM_007130.2	p.Met768Cysfs*48	A/AT	0.0308/ 0.0269/ 0.0294	A1A1=0/ A1R=1/ A1=2/ RR=3949/ R=2305	Yes	122	0.8	exon 5/5
X:46360032	ZNF674 ^a	NM_001039891.2	p.Ile331*	A/ATG	0.3847/ 0.1554/ 0.3067	A1A1=13/ A1R=1/ A1=2/ RR=3602/ R=2223	No	85	-0.4	exon 6/6
X:79958985	BRWD3	NM_153252.4	p.Asp944Argfs*2	TC/T	0.2006/ 0.1074/ 0.1666	A1A1=0/ A1R=14/ A1=3/ RR=3936/ R=2302	Yes	45	3.8	exon24/41. Insertion of a G after 3 Gs.
X:71694561	HDAC8 b	NM_001166419.1	p.Pro252Glnfs*18	T/TG	0.0/ 0.141/ 0.0481	A1A1=0/ A1R=4/ RR=3114/ R=2081	Yes	87	2.8	Alternative exon8/8 (2/6 isoformes). Del of a G in a stretch of 4 Gs.
X:53310691	IQSEC2 a, b	NM_015075.1	p.Arg6Glyfs*24	C/CG	0.2056/ 0.259/ 0.2252	A1A1=0/ A1R=15/ A1=4/ RR=3404/ R=1596	Yes	7	-6.6	Alternative exon 1/1. Del of a G, in a stretch of 6 Gs.
X:53279788	IQSEC2 ^a	NM_001111125.2	p.Ala657Profs*35	G/GCT	0.4293/ 1.0725/ 0.661	A1A1=8/ A1R=38/ A1=11/ RR=3763/ R=2204	Yes	15	4.5	exon 5/14-15
X:153296066	МЕСР2	NM_001110792.1	p.Glu416Alafs*31 (rs61752992)	G/GCT	0.0155/ 0.0/ 0.0098	A1A1=0/ A1R=1/ RR=3943/ R=2272	Yes	53	5.6	exon4/4.

X:70360675	MED12	NM_005120.2	p.Ile2079Serfs*139	A/ATCCG	0.7466/ 1.4497/ 1.0049	A1A1=1/ A1R=79/ A1=19/ RR=3805/ R=2162	Yes	21	4.4	exon 42/45. Exon polyQ-rich
X:70360677	MED12	NM_005120.2	p.Arg2080Asnfs*140	CAA/C	0.3812/ 0.5747/ 0.4523	A1A1=1/ A1R=36/ A1=7/ RR=3850/ R=2169	Yes	21	3.4	exon 42/45. Exon polyQ-rich
X:17750060	NHS	NM_001136024.2	p.Ser1457Lysfs*12	A/AGC	0.0154/ 0.0/ 0.0098	A1A1=0/ A1R=1/ RR=3949/ R=2307	Yes	132	2.4	exon 10/10. del GC in a stretch of AGC repeats, poly-S rich.
X:17750063	NHS	NM_001136024.2	p.Ser1458Thrfs*13	A/AG	0.0154/ 0.0269/ 0.0196	A1A1=0/ A1R=1/ A1=1 / RR=3949/ R=2306	Yes	131	2.8	exon 10/10. del of a G in a stretch of AGC repeats, poly-S rich.
X:101096513	NXF5 ^a	NM_032946.2	p.Ser86Ilefs*9	A/AC	0.0154/ 0.0/ 0.0098	A1A1=0/ A1R=1/ RR=3949/ R=2307	Yes	147	-0.3	exon 6/16.
X:139586517	SOX3	NM_005634.2	p.Ala236Glyfs*141	C/CGG	0.1446/ 0.1269/ 0.1382	A1A1=1/ A1R=5/ A1=5/ RR=3501/ R=1664	Yes	5	1.4	exon 1/1. In a region of CGG repeats, poly-Alanine rich.
X:117959888	ZCCHC12ª	NM_173798.2	p.Lys228Asnfs*27	T/TA	0.0154/ 0.0/ 0.0098	A1A1=0/ A1R=1/ RR=3949/ R=2307	Yes	55	1.8	exon 4/4. del of A, in 3 As.

X:70472962	ZMYM3	NM_001171162.1	p.Pro48Leufs*65	A/AG	0.5971/ 0.7236/ 0.6428	A1A1=7/ A1R=25/ A1=25/ RR=3826/ R=2216	No	13	0.6	exon 3/26. Del of a C, in a stretch of 7 Cs.
X:47307488	ZNF41 ^a	NM_007130.2	p.Phe560Leufs*35	T/TG	0.0154/ 0.0/ 0.0098	A1A1=0/ A1R=1/ RR=3949/ R=2307	Yes	74	1.5	exon 5/5
X:84526124	ZNF711 ^a	NM_021998.4	p.Thr526Lysfs*23	A/ACCCATACTG GTGAG	0.0463/ 0.0269/ 0.0392	A1A1=0/ A1R=3/ A1=1 / RR=3947/ R=2306	Yes	45	5.2	exon 9/9

^a: genes further discussed in this paper. ^b: the resulting frameshift affects only some specific isoforms. In red: frameshifts reported in hemizygous males, or homozygous females. HW: allelic frequencies in agreement with Hardy-Weinberg equilibrium.

Table S5. Statistical Analyses on the 104 Genes: Number of Non-synonymous (N) and Synonymous (S) Variants Reported in EVS for Each Gene and Subsequent dN/dS Ratios, Computed as Detailed in Material & Methods

Gene	Transcript -	EVS		N & S sites in mRNA			dN/dS ratios		
		N	S	N sites	S sites	protein length	dN	dS	ratio
ABCD1	NM_000033.3	25	28	1647	588	745	0,015	0,048	0,319
ACSL4/FACL4	NM_022977	20	14	1642	491	711	0,012	0,028	0,428
AFF2/FMR2	NM_002025.3	57	34	3027	906	1311	0,019	0,038	0,502
AGTR2 a	NM_000686.4	19	8	833	256	363	0,023	0,031	0,731 ^d
AP1S2	NM_003916.3	2	4	367	104	157	0,005	0,038	0,142
ARHGEF6 ^a	NM_004840.2	28	18	1801	528	776	0,016	0,034	0,456
ARHGEF9	NM_015185.2	12	13	1226	322	516	0,010	0,040	0,243
ARX	NM_139058.2	3	7	1236	451	562	0,002	0,016	0,156
ATP6AP2 ^a	NM_005765.2	14	10	797	253	350	0,018	0,040	0,444
ATP7A	NM_000052.4	70	31	3424	1077	1500	0,020	0,029	0,710
ATRX	NM_000489.3	61	44	5915	1561	2492	0,010	0,028	0,366
BCOR	NM_001123385.1	93	63	3998	1267	1755	0,023	0,050	0,468
BRWD3	NM_153252.4	33	33	4169	1238	1802	0,008	0,027	0,297
CASK	NM_003688.3	17	24	2131	632	921	0,008	0,038	0,210
CCDC22 b	NM_014008.3	40	17	1400	481	627	0,029	0,035	0,809 ^d
CDKL5	NM_003159.2	28	33	2375	715	1030	0,012	0,046	0,255
CLIC2 b	NM_001289.4	6	5	575	166	247	0,010	0,030	0,346
CNKSR2 b	NM_014927.3	22	19	2398	704	1034	0,009	0,027	0,340
CUL4B	NM_003588.3	13	12	2150	589	913	0,006	0,020	0,297
DCX	NM_000555, NM_178152	8	5	1019	304	441	0,008	0,016	0,478
DKC1	NM_001363.3	10	16	1184	358	514	0,008	0,045	0,189
DLG3	NM_021120.3	16	20	1863	588	817	0,009	0,034	0,253
DMD	NM_000109.3	224	114	8650	2382	3677	0,026	0,048	0,541
FANCB	NM_001018113.1	50	10	2013	564	859	0,025	0,018	1,400 ^d
FGD1	NM_004463.2	25	30	2167	716	961	0,012	0,042	0,276
FLNA	NM_001110556.1	97	125	5952	1989	2647	0,016	0,063	0,259
FMR1	NM_002024.5	16	14	1465	431	632	0,011	0,033	0,336
FRMPD4 b	NM_014728.3	57	46	3030	936	1322	0,019	0,049	0,383
FTSJ1	NM_177439.1	15	16	738	244	327	0,020	0,066	0,310
GDI1	NM_001493.2	8	20	1027	314	447	0,008	0,064	0,122
GK	NM_001205019.1	9	12	1288	390	559	0,007	0,031	0,227
GPC3	NM_001164617.1	25	15	1403	406	603	0,018	0,037	0,483
GRIA3	NM_007325.4	19	17	2061	622	894	0,009	0,027	0,337
HADH2	NM_004493.2	4	4	578	206	261	0,007	0,019	0,356
HCFC1 b	NM_005334.2	50	80	4477	1628	2035	0,011	0,049	0,227
HCCS	NM_005333.4	10	6	625	180	268	0,016	0,033	0,479
HDAC8	NM_018486.2	8	8	860	271	377	0,009	0,030	0,315
HPRT1	NM_000194.2	4	5	504	150	218	0,008	0,033	0,238
HUWE1 c	NM_031407.4	67	105	9976	3147	4374	0,007	0,033	0,201

IDS	NM_000202.5	38	20	1248	402	550	0,030	0,050	0,612
<i>IGBP1</i> b	NM_001551.2	19	11	793	224	339	0,024	0,049	0,487
IKBKG	NM_001099857.1	12	12	980	277	419	0,012	0,043	0,283
IL1RAPL1	NM_014271.3	14	22	1606	482	696	0,009	0,046	0,191
IQSEC2	NM_001111125.1	14	20	3334	1130	1488	0,004	0,018	0,237
KDM5C	NM_004187.3	35	42	3532	1148	1560	0,010	0,037	0,271
SHROOM4 b	NM_020717.3	86	28	3448	1031	1493	0,025	0,027	0,918 ^d
<i>KIAA2022</i> b	NM_001008537.2	54	34	3559	989	1516	0,015	0,034	0,441
KLF8 b	NM_007250.4	16	6	821	256	359	0,019	0,023	0,831 ^d
LICAM	NM_000425.3	53	48	2866	905	1257	0,018	0,053	0,348
LAMP2	NM_002294.2	25	12	933	297	410	0,027	0,040	0,663
MAGT1/IAP ^a	NM_032121.5	15	10	846	255	367	0,018	0,039	0,453
MAOA b	NM_000240.3	14	12	1209	372	527	0,012	0,032	0,359
MBTPS2	NM_015884.3	21	13	1173	384	519	0,018	0,034	0,529
MECP2	NM_004992.3	37	45	1102	356	486	0,034	0,126	0,266
MED12	NM_005120.2	26	50	4982	1549	2177	0,005	0,032	0,162
MID1	NM_000381.3	25	20	1539	462	667	0,016	0,043	0,375
NAA10 b	NM_003491.2	2	8	544	161	235	0,004	0,050	0,074
NDP	NM_000266.3	7	0	302	97	133	0,023	0,000	-
NDUFA1	NM_004541.3	4	3	161	49	70	0,025	0,062	0,402
NHS	NM_198270.2	73	37	3711	1179	1630	0,020	0,031	0,627
NLGN3 b	NM_181303.1	17	27	1910	634	848	0,009	0,043	0,209
NLGN4X	NM_020742.2	20	40	1868	580	816	0,011	0,069	0,155
NSDHL	NM_015922.2	34	17	851	268	373	0,040	0,064	0,629
NXF5 ^a	NM_032946.2	30	11	851	244	365	0,035	0,045	0,783 ^d
OCRL	NM_000276.3	27	15	2105	598	901	0,013	0,025	0,511
OFD1	NM_003611.2	49	22	2381	655	1012	0,021	0,034	0,613
OPHN1	NM_002547.2	34	18	1866	540	802	0,018	0,033	0,547
ОТС	NM_000531.5	17	10	815	247	354	0,021	0,040	0,515
PAK3	NM_001128168.1	13	14	1342	398	580	0,010	0,035	0,275
PCDH19	NM_001105243.1	37	36	2492	812	1101	0,015	0,044	0,335
PDHA1	NM_000284.3	15	8	890	280	390	0,017	0,029	0,589
PGK1	NM_000291.3	24	5	959	293	417	0,025	0,017	1,465 ^d
PHF6	NM_001015877.1	6	4	859	236	365	0,007	0,017	0,411
PHF8	NM_001184896.1	17	26	2428	752	1060	0,007	0,035	0,203
PLP1	NM_001128834.1	2	6	620	211	277	0,003	0,028	0,113
PORCN	NM_203475.1	16	16	1029	355	461	0,016	0,045	0,345
PQBP1	NM_005710.2	2	8	606	189	265	0,003	0,042	0,078
PRPS1	NM_002764.3	0	7	734	220	318	0,000	0,032	0,000
PTCHD1 ^c	NM_173495.2	29	25	2025	639	888	0,014	0,039	0,366
RAB39B	NM_171998.2	1	7	488	151	213	0,002	0,046	0,044
RBM10	NM_001204468.1	16	35	2263	722	995	0,007	0,048	0,146
RPL10 b	NM_006013.3	2	4	491	151	214	0,004	0,027	0,154
RPS6KA3	NM_004586.2	10	14	1712	508	740	0,006	0,028	0,212
SLC16A2	NM_006517.3	20	9	1218	400	539	0,016	0,023	0,729 ^d

SLC6A8	M_005629.3	14	29	1426	479	635	0,010	0,061	0,162
SLC9A6	NM_001042537.1	11	16	1592	512	701	0,007	0,031	0,221
SMC1A	NM_006306.2	8	23	2911	788	1233	0,003	0,029	0,094
SMS	NM_004595.3	6	16	846	252	366	0,007	0,064	0,112
SOX3	NM_005634.2	6	8	977	361	446	0,006	0,022	0,277
SRPX2 ^a	NM_014467.2	31	14	1050	345	465	0,030	0,041	0,728 ^d
SYN1 ^c	NM_006950.3	12	12	1570	545	705	0,008	0,022	0,347
SYP	NM_003179.2	9	8	705	234	313	0,013	0,034	0,373
TIMM8A	NM_004085.3	3	4	230	62	97	0,013	0,065	0,201
TSPAN7	NM_004615.3	9	3	564	183	249	0,016	0,016	0,973 ^d
UBE2A	NM_003336.2	0	1	351	105	152	0,000	0,010	0,000
UPF3B	NM_080632.2	19	10	1153	297	483	0,016	0,034	0,489
ZCCHC12/ SIZN1 a	NM_173798.2	19	14	928	278	402	0,020	0,050	0,406
ZDHHC9	NM_016032.3	2	12	832	260	364	0,002	0,046	0,052
ZDHHC15 ^b	NM_144969.2	17	8	781	231	337	0,022	0,035	0,628
ZNF261/ZMYM3 ^b	NM_201599.2	31	42	3129	982	1370	0,010	0,043	0,232
ZNF41 ^a	NM_007130.2	45	17	1850	487	779	0,024	0,035	0,696
ZNF674 ^a	NM_001039891.2	28	10	1375	368	581	0,020	0,027	0,750 ^d
ZNF711	NM_021998.4	15	15	1795	489	761	0,008	0,031	0,272
ZNF81 ^a	NM_007137.3	25	11	1572	411	661	0,016	0,027	0,595

^{a, b, c}: genes further discussed in this paper; ^a: Gene for which implication in ID is questioned regarding current evidences and EVS data; ^b: genes missing replication studies in order to be fully considered as implicated in ID; ^c: genes for which implication in ID is very likely, despite some contradictory observations in EVS. ^d: dS/dN ratios superior to the 90th percentile (0.723)

References

- Kleefstra T, Yntema HG, Oudakker AR, Banning MJ, Kalscheuer VM, Chelly J, Moraine C, Ropers HH, Fryns JP, Janssen IM, Sistermans EA, Nillesen WN, de Vries LB, Hamel BC, van Bokhoven H (2004) Zinc finger 81 (ZNF81) mutations associated with X-linked mental retardation. J Med Genet 41:394-399
- 2. Tarpey PS, Smith R, Pleasance E, Whibley A, Edkins S, Hardy C, O'Meara S, et al. (2009) A systematic, large-scale resequencing screen of X-chromosome coding exons in mental retardation. Nat Genet 41:535-543
- 3. Alesi V, Bertoli M, Barrano G, Torres B, Pusceddu S, Pastorino M, Perria C, Nardone AM, Novelli A, Serra G (2012) 335.4 kb microduplication in chromosome band Xp11.2p11.3 associated with developmental delay, growth retardation, autistic disorder and dysmorphic features. Gene 505:384-387
- 4. El-Hattab AW, Bournat J, Eng PA, Wu JB, Walker BA, Stankiewicz P, Cheung SW, Brown CW (2011) Microduplication of Xp11.23p11.3 with effects on cognition, behavior, and craniofacial development. Clin Genet 79:531-538
- 5. Ramser J, Abidi FE, Burckle CA, Lenski C, Toriello H, Wen G, Lubs HA, Engert S, Stevenson RE, Meindl A, Schwartz CE, Nguyen G (2005) A unique exonic splice enhancer mutation in a family with X-linked mental retardation and epilepsy points to a novel role of the renin receptor. Hum Mol Genet 14:1019-1027
- 6. Cho G, Bhat SS, Gao J, Collins JS, Rogers RC, Simensen RJ, Schwartz CE, Golden JA, Srivastava AK (2008) Evidence that SIZN1 is a candidate X-linked mental retardation gene. Am J Med Genet A 146A:2644-2650
- 7. Graham JM, Jr., Wheeler P, Tackels-Horne D, Lin AE, Hall BD, May M, Short KM, Schwartz CE, Cox TC (2003) A new X-linked syndrome with agenesis of the corpus callosum, mental retardation, coloboma, micrognathia, and a mutation in the Alpha 4 gene at Xq13. Am J Med Genet A 123A:37-44
- 8. Jamain S, Quach H, Betancur C, Rastam M, Colineaux C, Gillberg IC, Soderstrom H, Giros B, Leboyer M, Gillberg C, Bourgeron T (2003) Mutations of the X-linked genes encoding neuroligins NLGN3 and NLGN4 are associated with autism. Nat Genet 34:27-29
- 9. Chih B, Afridi SK, Clark L, Scheiffele P (2004) Disorder-associated mutations lead to functional inactivation of neuroligins. Hum Mol Genet 13:1471-1477
- 10. Tabuchi K, Blundell J, Etherton MR, Hammer RE, Liu X, Powell CM, Sudhof TC (2007) A neuroligin-3 mutation implicated in autism increases inhibitory synaptic transmission in mice. Science 318:71-76
- 11. Talebizadeh Z, Lam DY, Theodoro MF, Bittel DC, Lushington GH, Butler MG (2006) Novel splice isoforms for NLGN3 and NLGN4 with possible implications in autism. J Med Genet 43:e21
- 12. Daoud H, Bonnet-Brilhault F, Vedrine S, Demattei MV, Vourc'h P, Bayou N, Andres CR, Barthelemy C, Laumonnier F, Briault S (2009) Autism and nonsyndromic mental retardation associated with a de novo mutation in the NLGN4X gene promoter causing an increased expression level. Biol Psychiatry 66:906-910
- 13. Laumonnier F, Bonnet-Brilhault F, Gomot M, Blanc R, David A, Moizard MP, Raynaud M, Ronce N, Lemonnier E, Calvas P, Laudier B, Chelly J, Fryns JP, Ropers HH, Hamel BC, Andres C, Barthelemy C, Moraine C, Briault S (2004) X-linked mental retardation and autism are associated with a mutation in the NLGN4 gene, a member of the neuroligin family. Am J Hum Genet 74:552-557
- 14. Lawson-Yuen A, Saldivar JS, Sommer S, Picker J (2008) Familial deletion within NLGN4 associated with autism and Tourette syndrome. Eur J Hum Genet 16:614-618
- 15. Blasi F, Bacchelli E, Pesaresi G, Carone S, Bailey AJ, Maestrini E (2006) Absence of coding mutations in the X-linked genes neuroligin 3 and neuroligin 4 in individuals with autism from the IMGSAC collection. Am J Med Genet B Neuropsychiatr Genet 141B:220-221

- 16. Gauthier J, Bonnel A, St-Onge J, Karemera L, Laurent S, Mottron L, Fombonne E, Joober R, Rouleau GA (2005) NLGN3/NLGN4 gene mutations are not responsible for autism in the Quebec population. Am J Med Genet B Neuropsychiatr Genet 132B:74-75
- 17. Steinberg KM, Ramachandran D, Patel VC, Shetty AC, Cutler DJ, Zwick ME (2012) Identification of rare X-linked neuroligin variants by massively parallel sequencing in males with autism spectrum disorder. Mol Autism 3:8
- 18. Vincent JB, Kolozsvari D, Roberts WS, Bolton PF, Gurling HM, Scherer SW (2004) Mutation screening of X-chromosomal neuroligin genes: no mutations in 196 autism probands. Am J Med Genet B Neuropsychiatr Genet 129B:82-84
- 19. Wermter AK, Kamp-Becker I, Strauch K, Schulte-Korne G, Remschmidt H (2008) No evidence for involvement of genetic variants in the X-linked neuroligin genes NLGN3 and NLGN4X in probands with autism spectrum disorder on high functioning level. Am J Med Genet B Neuropsychiatr Genet 147B:535-537
- 20. Ylisaukko-oja T, Rehnstrom K, Auranen M, Vanhala R, Alen R, Kempas E, Ellonen P, Turunen JA, Makkonen I, Riikonen R, Nieminen-von Wendt T, von Wendt L, Peltonen L, Jarvela I (2005) Analysis of four neuroligin genes as candidates for autism. Eur J Hum Genet 13:1285-1292
- 21. Voineagu I, Huang L, Winden K, Lazaro M, Haan E, Nelson J, McGaughran J, Nguyen LS, Friend K, Hackett A, Field M, Gecz J, Geschwind D (2012) CCDC22: a novel candidate gene for syndromic X-linked intellectual disability. Mol Psychiatry 17:4-7
- 22. Witham S, Takano K, Schwartz C, Alexov E (2011) A missense mutation in CLIC2 associated with intellectual disability is predicted by in silico modeling to affect protein stability and dynamics. Proteins 79:2444-2454
- 23. Takano K, Liu D, Tarpey P, Gallant E, Lam A, Witham S, Alexov E, Chaubey A, Stevenson RE, Schwartz CE, Board PG, Dulhunty AF (2012) An X-linked channelopathy with cardiomegaly due to a CLIC2 mutation enhancing ryanodine receptor channel activity. Hum Mol Genet 21:4497-4507
- 24. Huang L, Jolly LA, Willis-Owen S, Gardner A, Kumar R, Douglas E, Shoubridge C, Wieczorek D, Tzschach A, Cohen M, Hackett A, Field M, Froyen G, Hu H, Haas SA, Ropers HH, Kalscheuer VM, Corbett MA, Gecz J (2012) A noncoding, regulatory mutation implicates HCFC1 in nonsyndromic intellectual disability. Am J Hum Genet 91:694-702
- 25. Piton A, Gauthier J, Hamdan FF, Lafreniere RG, Yang Y, Henrion E, Laurent S, et al. (2011) Systematic resequencing of X-chromosome synaptic genes in autism spectrum disorder and schizophrenia. Mol Psychiatry 16:867-880
- 26. Rope AF, Wang K, Evjenth R, Xing J, Johnston JJ, Swensen JJ, Johnson WE, et al. (2011) Using VAAST to identify an X-linked disorder resulting in lethality in male infants due to N-terminal acetyltransferase deficiency. Am J Hum Genet 89:28-43
- 27. Rauch A, Wieczorek D, Graf E, Wieland T, Endele S, Schwarzmayr T, Albrecht B, et al. (2012) Range of genetic mutations associated with severe non-syndromic sporadic intellectual disability: an exome sequencing study. Lancet 380:1674-1682
- 28. Chiocchetti A, Pakalapati G, Duketis E, Wiemann S, Poustka A, Poustka F, Klauck SM (2011) Mutation and expression analyses of the ribosomal protein gene RPL10 in an extended German sample of patients with autism spectrum disorder. Am J Med Genet A 155A:1472-1475
- 29. Klauck SM, Felder B, Kolb-Kokocinski A, Schuster C, Chiocchetti A, Schupp I, Wellenreuther R, Schmotzer G, Poustka F, Breitenbach-Koller L, Poustka A (2006) Mutations in the ribosomal protein gene RPL10 suggest a novel modulating disease mechanism for autism. Mol Psychiatry 11:1073-1084
- 30. Gong X, Delorme R, Fauchereau F, Durand CM, Chaste P, Betancur C, Goubran-Botros H, Nygren G, Anckarsater H, Rastam M, Gillberg IC, Kopp S, Mouren-Simeoni MC, Gillberg C, Leboyer M, Bourgeron T (2009) An investigation of ribosomal protein L10 gene in autism spectrum disorders. BMC Med Genet 10:7

- 31. De Keersmaecker K, Atak ZK, Li N, Vicente C, Patchett S, Girardi T, Gianfelici V, et al. (2012) Exome sequencing identifies mutation in CNOT3 and ribosomal genes RPL5 and RPL10 in T-cell acute lymphoblastic leukemia. Nat Genet
- 32. Hagens O, Dubos A, Abidi F, Barbi G, Van Zutven L, Hoeltzenbein M, Tommerup N, Moraine C, Fryns JP, Chelly J, van Bokhoven H, Gecz J, Dollfus H, Ropers HH, Schwartz CE, de Cassia Stocco Dos Santos R, Kalscheuer V, Hanauer A (2006) Disruptions of the novel KIAA1202 gene are associated with X-linked mental retardation. Hum Genet 118:578-590
- 33. Honda S, Hayashi S, Imoto I, Toyama J, Okazawa H, Nakagawa E, Goto Y, Inazawa J (2010) Copynumber variations on the X chromosome in Japanese patients with mental retardation detected by array-based comparative genomic hybridization analysis. J Hum Genet 55:590-599
- 34. Cantagrel V, Lossi AM, Boulanger S, Depetris D, Mattei MG, Gecz J, Schwartz CE, Van Maldergem L, Villard L (2004) Disruption of a new X linked gene highly expressed in brain in a family with two mentally retarded males. J Med Genet 41:736-742
- 35. Lossi AM, Laugier-Anfossi F, Depetris D, Gecz J, Gedeon A, Kooy F, Schwartz C, Mattei MG, Croquette MF, Villard L (2002) Abnormal expression of the KLF8 (ZNF741) gene in a female patient with an X;autosome translocation t(X;21)(p11.2;q22.3) and non-syndromic mental retardation. J Med Genet 39:113-117
- 36. Mansouri MR, Marklund L, Gustavsson P, Davey E, Carlsson B, Larsson C, White I, Gustavson KH, Dahl N (2005) Loss of ZDHHC15 expression in a woman with a balanced translocation t(X;15)(q13.3;cen) and severe mental retardation. Eur J Hum Genet 13:970-977
- 37. van der Maarel SM, Scholten IH, Huber I, Philippe C, Suijkerbuijk RF, Gilgenkrantz S, Kere J, Cremers FP, Ropers HH (1996) Cloning and characterization of DXS6673E, a candidate gene for X-linked mental retardation in Xq13.1. Hum Mol Genet 5:887-897
- 38. Honda S, Orii KO, Kobayashi J, Hayashi S, Imamura A, Imoto I, Nakagawa E, Goto Y, Inazawa J (2010) Novel deletion at Xq24 including the UBE2A gene in a patient with X-linked mental retardation. J Hum Genet 55:244-247
- 39. Houge G, Rasmussen IH, Hovland R (2012) Loss-of-Function CNKSR2 Mutation Is a Likely Cause of Non-Syndromic X-Linked Intellectual Disability. Mol Syndromol 2:60-63
- 40. Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J, Shago M, et al. (2008) Structural variation of chromosomes in autism spectrum disorder. Am J Hum Genet 82:477-488
- 41. Pinto D, Pagnamenta AT, Klei L, Anney R, Merico D, Regan R, Conroy J, et al. (2010) Functional impact of global rare copy number variation in autism spectrum disorders. Nature 466:368-372
- 42. Whibley AC, Plagnol V, Tarpey PS, Abidi F, Fullston T, Choma MK, Boucher CA, Shepherd L, Willatt L, Parkin G, Smith R, Futreal PA, Shaw M, Boyle J, Licata A, Skinner C, Stevenson RE, Turner G, Field M, Hackett A, Schwartz CE, Gecz J, Stratton MR, Raymond FL (2010) Fine-scale survey of X chromosome copy number variants and indels underlying intellectual disability. Am J Hum Genet 87:173-188
- 43. Filges I, Rothlisberger B, Blattner A, Boesch N, Demougin P, Wenzel F, Huber AR, Heinimann K, Weber P, Miny P (2010) Deletion in Xp22.11: PTCHD1 is a candidate gene for X-linked intellectual disability with or without autism. Clin Genet 79:79-85
- 44. Noor A, Whibley A, Marshall CR, Gianakopoulos PJ, Piton A, Carson AR, Orlic-Milacic M, et al. (2010) Disruption at the PTCHD1 Locus on Xp22.11 in Autism spectrum disorder and intellectual disability. Sci Transl Med 2:49ra68
- 45. DeGennaro LJ, McCaffery CA, Kirchgessner CU, Yang-Feng TL, Francke U (1987) Molecular analysis of synapsin I, a candidate gene for Rett syndrome. Brain Dev 9:469-474
- 46. Ferlini A, Ansaloni L, Nobile C, Forabosco A (1990) Molecular analysis of the Rett syndrome using cDNA synapsin I as a probe. Brain Dev 12:136-139
- 47. Garcia CC, Blair HJ, Seager M, Coulthard A, Tennant S, Buddles M, Curtis A, Goodship JA (2004) Identification of a mutation in synapsin I, a synaptic vesicle protein, in a family with epilepsy. J Med Genet 41:183-186

- 48. Fassio A, Patry L, Congia S, Onofri F, Piton A, Gauthier J, Pozzi D, Messa M, Defranchi E, Fadda M, Corradi A, Baldelli P, Lapointe L, St-Onge J, Meloche C, Mottron L, Valtorta F, Khoa Nguyen D, Rouleau GA, Benfenati F, Cossette P (2011) SYN1 loss-of-function mutations in autism and partial epilepsy cause impaired synaptic function. Hum Mol Genet 20:2297-2307
- 49. Li L, Chin LS, Shupliakov O, Brodin L, Sihra TS, Hvalby O, Jensen V, Zheng D, McNamara JO, Greengard P, et al. (1995) Impairment of synaptic vesicle clustering and of synaptic transmission, and increased seizure propensity, in synapsin I-deficient mice. Proc Natl Acad Sci U S A 92:9235-9239
- 50. Takei Y, Harada A, Takeda S, Kobayashi K, Terada S, Noda T, Takahashi T, Hirokawa N (1995) Synapsin I deficiency results in the structural change in the presynaptic terminals in the murine nervous system. J Cell Biol 131:1789-1800
- 51. Corradi A, Zanardi A, Giacomini C, Onofri F, Valtorta F, Zoli M, Benfenati F (2008) Synapsin-I- and synapsin-II-null mice display an increased age-dependent cognitive impairment. J Cell Sci 121:3042-3051
- 52. Lubs HA, Stevenson RE, Schwartz CE (2012) Fragile X and X-linked intellectual disability: four decades of discovery. Am J Hum Genet 90:579-590
- 53. Ropers HH (2010) Genetics of early onset cognitive impairment. Annu Rev Genomics Hum Genet 11:161-187
- 54. Vervoort VS, Beachem MA, Edwards PS, Ladd S, Miller KE, de Mollerat X, Clarkson K, DuPont B, Schwartz CE, Stevenson RE, Boyd E, Srivastava AK (2002) AGTR2 mutations in X-linked mental retardation. Science 296:2401-2403
- 55. Ylisaukko-oja T, Rehnstrom K, Vanhala R, Tengstrom C, Lahdetie J, Jarvela I (2004) Identification of two AGTR2 mutations in male patients with non-syndromic mental retardation. Hum Genet 114:211-213
- 56. Erdmann J, Dahmlow S, Guse M, Hetzer R, Regitz-Zagrosek V (2004) The assertion that a G21V mutation in AGTR2 causes mental retardation is not supported by other studies. Hum Genet 114:396; author reply 397
- 57. Kutsche K, Yntema H, Brandt A, Jantke I, Nothwang HG, Orth U, Boavida MG, David D, Chelly J, Fryns JP, Moraine C, Ropers HH, Hamel BC, van Bokhoven H, Gal A (2000) Mutations in ARHGEF6, encoding a guanine nucleotide exchange factor for Rho GTPases, in patients with X-linked mental retardation. Nat Genet 26:247-250
- 58. Lenk U, Hanke R, Speer A (1994) Carrier detection in DMD families with point mutations, using PCR-SSCP and direct sequencing. Neuromuscul Disord 4:411-418
- 59. Lebel RR, May M, Pouls S, Lubs HA, Stevenson RE, Schwartz CE (2002) Non-syndromic X-linked mental retardation associated with a missense mutation (P312L) in the FGD1 gene. Clin Genet 61:139-145
- 60. Hehr U, Hehr A, Uyanik G, Phelan E, Winkler J, Reardon W (2006) A filamin A splice mutation resulting in a syndrome of facial dysmorphism, periventricular nodular heterotopia, and severe constipation reminiscent of cerebro-fronto-facial syndrome. J Med Genet 43:541-544
- 61. Unger S, Mainberger A, Spitz C, Bahr A, Zeschnigk C, Zabel B, Superti-Furga A, Morris-Rosendahl DJ (2007) Filamin A mutation is one cause of FG syndrome. Am J Med Genet A 143A:1876-1879
- 62. Liang GS, de Miguel M, Gomez-Hernandez JM, Glass JD, Scherer SS, Mintz M, Barrio LC, Fischbeck KH (2005) Severe neuropathy with leaky connexin32 hemichannels. Ann Neurol 57:749-754
- 63. Molinari F, Foulquier F, Tarpey PS, Morelle W, Boissel S, Teague J, Edkins S, Futreal PA, Stratton MR, Turner G, Matthijs G, Gecz J, Munnich A, Colleaux L (2008) Oligosaccharyltransferase-subunit mutations in nonsyndromic mental retardation. Am J Hum Genet 82:1150-1157
- 64. Potluri P, Davila A, Ruiz-Pesini E, Mishmar D, O'Hearn S, Hancock S, Simon M, Scheffler IE, Wallace DC, Procaccio V (2009) A novel NDUFA1 mutation leads to a progressive mitochondrial complex I-specific neurodegenerative disease. Mol Genet Metab 96:189-195

- 65. Matthews PM, Brown RM, Otero LJ, Marchington DR, LeGris M, Howes R, Meadows LS, Shevell M, Scriver CR, Brown GK (1994) Pyruvate dehydrogenase deficiency. Clinical presentation and molecular genetic characterization of five new patients. Brain 117 (Pt 3):435-443
- 66. Roll P, Rudolf G, Pereira S, Royer B, Scheffer IE, Massacrier A, Valenti MP, et al. (2006) SRPX2 mutations in disorders of language cortex and cognition. Hum Mol Genet 15:1195-1207
- 67. Gomot M, Ronce N, Dessay S, Zemni R, Ayrault AD, Moizard MP, Nivelon A, Gilgenkrantz S, Dourlens J, Des Portes V, Chelly J, Moraine C (2002) TM4SF2 gene involvement reconsidered in an XLMR family after neuropsychological assessment. Am J Med Genet 112:400-404
- 68. Maranduba CM, Sa Moreira E, Muller Orabona G, Pavanello RC, Vianna-Morgante AM, Passos-Bueno MR (2004) Does the P172H mutation at the TM4SF2 gene cause X-linked mental retardation? Am J Med Genet A 124A:413-415
- 69. Zemni R, Bienvenu T, Vinet MC, Sefiani A, Carrie A, Billuart P, McDonell N, et al. (2000) A new gene involved in X-linked mental retardation identified by analysis of an X;2 balanced translocation. Nat Genet 24:167-170
- 70. Shoichet SA, Hoffmann K, Menzel C, Trautmann U, Moser B, Hoeltzenbein M, Echenne B, Partington M, Van Bokhoven H, Moraine C, Fryns JP, Chelly J, Rott HD, Ropers HH, Kalscheuer VM (2003) Mutations in the ZNF41 gene are associated with cognitive deficits: identification of a new candidate for X-linked mental retardation. Am J Hum Genet 73:1341-1354